

2-28-2023

Chromosome-Length Assembly of the Baikal Seal (*Pusa sibirica*) Genome Reveals a Historically Large Population Prior to Isolation in Lake Baikal

Aliya Yakupova
ITMO University

Andrey Tomarovsky
ITMO University; Novosibirsk State University; Institute of Molecular and Cellular Biology SB RAS

Azamat Totikov
ITMO University; Novosibirsk State University; Institute of Molecular and Cellular Biology SB RAS

Violetta R. Beklemisheva
Institute of Molecular and Cellular Biology SB RAS

Maria Logacheva
Skolkovo Institute of Science and Technology

Follow this and additional works at: https://nsuworks.nova.edu/cnso_bio_facarticles



Part of the [Biology Commons](#)
See next page for additional authors

NSUWorks Citation

Yakupova, Aliya; Andrey Tomarovsky; Azamat Totikov; Violetta R. Beklemisheva; Maria Logacheva; Polina Perelman; Aleksey Komissarov; Pavel Dobrynin; Ksenia Krasheninnikova; Gaik Tamazian; Natalya A. Serdyukova; Michael Rayko; Tatiana Bulyonkova; Nikolay Cherkasov; Vladimir Pylev; Vladimir Peterfeld; Aleksey Penin; Elena Balanovska; Alla L. Lapidus; DNA Zoo Consortium; Stephen James O'Brien; Alexander S. Graphodatsky; Klaus-Peter Koepfli; and Sergey Kliver. 2023. "Chromosome-Length Assembly of the Baikal Seal (*Pusa sibirica*) Genome Reveals a Historically Large Population Prior to Isolation in Lake Baikal." *Genes* 14, (3): 619. doi:10.3390/genes14030619.













This Article is brought to you for free and open access by the Department of Biological Sciences at NSUWorks. It has been accepted for inclusion in Biology Faculty Articles by an authorized administrator of NSUWorks. For more information, please contact nsuworks@nova.edu.

Authors

Aliya Yakupova, Andrey Tomarovsky, Azamat Totikov, Violetta R. Beklemisheva, Maria Logacheva, Polina Perelman, Aleksey Komissarov, Pavel Dobrynin, Ksenia Krasheninnikova, Gaik Tamazian, Natalya A. Serdyukova, Michael Rayko, Tatiana Bulyonkova, Nikolay Cherkasov, Vladimir Pylev, Vladimir Peterfeld, Aleksey Penin, Elena Balanovska, Alla L. Lapidus, DNA Zoo Consortium, Stephen James O'Brien, Alexander S. Graphodatsky, Klaus-Peter Koepfli, and Sergey Kliver

Article

Chromosome-Length Assembly of the Baikal Seal (*Pusa sibirica*) Genome Reveals a Historically Large Population Prior to Isolation in Lake Baikal

Aliya Yakupova ^{1,*} , Andrey Tomarovsky ^{1,2,3} , Azamat Totikov ^{1,2,3} , Violetta Beklemisheva ³ , Maria Logacheva ⁴, Polina L. Perelman ³ , Aleksey Komissarov ⁵ , Pavel Dobrynin ^{1,6}, Ksenia Krasheninnikova ⁷, Gaik Tamazian ⁸ , Natalia A. Serdyukova ³, Mike Rayko ⁹, Tatiana Bulyonkova ¹⁰, Nikolay Cherkasov ⁸, Vladimir Pylev ¹¹ , Vladimir Peterfeld ¹², Aleksey Penin ¹³, Elena Balanovska ¹¹ , Alla Lapidus ⁹ , DNA Zoo Consortium ¹⁴, Stephen J. OBrien ¹⁵, Alexander Graphodatsky ^{3,*} , Klaus-Peter Koepfli ^{16,17} , and Sergei Kliver ¹⁸

- ¹ Computer Technologies Laboratory, ITMO University, 19701 Saint Petersburg, Russia
 - ² Department of Natural Sciences, Novosibirsk State University, 630090 Novosibirsk, Russia
 - ³ Department of the Diversity and Evolution of Genomes, Institute of Molecular and Cellular Biology SB RAS, 630090 Novosibirsk, Russia
 - ⁴ Skolkovo Institute of Science and Technology, 121205 Moscow, Russia
 - ⁵ Applied Genomics Laboratory, SCAMT Institute, ITMO University, 9 Ulitsa Lomonosova, 191002 Saint Petersburg, Russia
 - ⁶ Human Genetics Laboratory, Vavilov Institute of General Genetics RAS, 119991 Moscow, Russia
 - ⁷ Tree of Life, Wellcome Sanger Institute, Cambridge CB10 1SA, UK
 - ⁸ Centre for Computational Biology, Peter the Great Saint Petersburg Polytechnic University, 195251 St. Petersburg, Russia
 - ⁹ Center for Bioinformatics and Algorithmic Biotechnology, St. Petersburg State University, 199034 St. Petersburg, Russia
 - ¹⁰ Laboratory of Mixed Computations, A.P. Ershov Institute of Informatics Systems SB RAS, 630090 Novosibirsk, Russia
 - ¹¹ Laboratory of Human Population Genetics, Research Centre for Medical Genetics, 115522 Moscow, Russia
 - ¹² Baikal Branch of State Research and Industrial Center of Fisheries, 670034 Ulan-Ude, Russia
 - ¹³ Institute for Information Transmission Problems of the Russian Academy of Sciences, 127051 Moscow, Russia
 - ¹⁴ The Center for Genome Architecture, Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, TX 77030, USA
 - ¹⁵ Guy Harvey Oceanographic Center, Halmos College of Arts and Sciences, NOVA Southeastern University, Fort Lauderdale, FL 33004, USA
 - ¹⁶ Smithsonian-Mason School of Conservation, George Mason University, 1500 Remount Road, Front Royal, VA 22630, USA
 - ¹⁷ Center for Species Survival, Smithsonian's National Zoo and Conservation Biology Institute, 1500 Remount Road, Front Royal, VA 22630, USA
 - ¹⁸ Center for Evolutionary Hologenomics, The Globe Institute, The University of Copenhagen, 5A, Oester Farimagsgade, 1353 Copenhagen, Denmark
- * Correspondence: aliyah.yakupova@gmail.com (A.Y.); graf@mcb.nsc.ru (A.G.)



Citation: Yakupova, A.; Tomarovsky, A.; Totikov, A.; Beklemisheva, V.; Logacheva, M.; Perelman, P.L.; Komissarov, A.; Dobrynin, P.; Krasheninnikova, K.; Tamazian, G.; et al. Chromosome-Length Assembly of the Baikal Seal (*Pusa sibirica*) Genome Reveals a Historically Large Population Prior to Isolation in Lake Baikal. *Genes* **2023**, *14*, 619. <https://doi.org/10.3390/genes14030619>

Academic Editor: Manuel Vera Rodriguez

Received: 21 December 2022

Revised: 31 January 2023

Accepted: 24 February 2023

Published: 28 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: *Pusa sibirica*, the Baikal seal, is the only extant, exclusively freshwater, pinniped species. The pending issue is, how and when they reached their current habitat—the rift lake Baikal, more than three thousand kilometers away from the Arctic Ocean. To explore the demographic history and genetic diversity of this species, we generated a de novo chromosome-length assembly, and compared it with three closely related marine pinniped species. Multiple whole genome alignment of the four species compared with their karyotypes showed high conservation of chromosomal features, except for three large inversions on chromosome VI. We found the mean heterozygosity of the studied Baikal seal individuals was relatively low (0.61 SNPs/kbp), but comparable to other analyzed pinniped samples. Demographic reconstruction of seals revealed differing trajectories, yet remarkable variations in N_e occurred during approximately the same time periods. The Baikal seal showed a significantly more severe decline relative to other species. This could be due to the difference in environmental conditions encountered by the earlier populations of Baikal seals, as ice sheets changed during glacial–interglacial cycles. We connect this period to the time of migration to

Lake Baikal, which occurred ~3–0.3 Mya, after which the population stabilized, indicating balanced habitat conditions.

Keywords: *Pusa sibirica*; conservation; pinnipeds; demography; heterozygosity

1. Introduction

The origin of the endemic freshwater Baikal seal (*Pusa sibirica*), and its phylogenetic relationships with other seals, is a long-term biogeographic issue. Lake Baikal is connected to the Kara Sea in the Arctic Ocean through the Angara–Yenisei river system. There is no definitive data of how seals migrated to a landlocked habitat, but there are several hypotheses. The most plausible current view suggests that their migration routes led from the Arctic waters along the Yenisei, or through glacial and ice-dammed lakes, which existed in the Middle Pleistocene during at least four continental glaciation periods in central Siberia [1,2]. Estimates of the migration time, inferred from phylogenetic analyses (primarily mitochondrial DNA), encompass a broad period, depending on the data and methods used: from 2–3 million years ago (Mya) (based on MP, ML, and NJ methods) [2], to 1.5–2 Mya (based on BI) [3], and 0.4 Mya (based on UPGMA and NJ methods) [4]. At the moment, researchers are sure only of the Arctic origin of the Baikal seal [5]. This hypothesis is supported by several Arctic adaptations of the modern pagophilic species: breeding on ice; the white natal fur of pups, providing camouflage on snow-covered ice; the ability to spend considerable time and cover long distances underwater, considering the chance to be trapped under the thick ice; and the instinct to breathe from pockets of exhaled air that accumulates under the ice [6–8]. Similar behavior is typical for the Baikal seal’s Arctic relatives—the ringed seal (*Pusa hispida*) and Caspian seal (*Pusa caspica*) [9–11].

The earliest records of seals in Lake Baikal appeared due to their interaction with humans. Hunter-fishermen began depicting seals on rocks, and as bone and stone carvings, more than 7000 years ago [7]. Zooarchaeological analyses of seal fossils showed that seal hunting began at least 9000 years ago, in the Middle Holocene [8]. Harvesting seals for their fat and fur began in the 19th and 20th centuries, and later, it continued in order to protect commercially important fish species. Poaching pressure became several times greater than that from legal harvesting, which became heavily regulated following a 1986 act by the Soviet government on the protection of marine mammals, that was caused by a global ban on commercial whaling, including all marine mammals [12]. The establishment of several federal nature protection territories along the Lake Baikal coastline, including the Zabaikalskiy and the Pribaikalskiy nature reserves (zakazniks), further contributed to conservation efforts. It was later revealed that, contrary to popular belief, the basis of the seal’s diet does not include commercially important fish, such as the omul (*Coregonus migratorius*), and does not pose a threat to fisheries [13,14]. Since Baikal seals lack molars for chewing, they prefer medium-sized fish, such as the Baikal oilfish, or golomyanka (genus *Comephorus*), which are swallowed whole. Baikal seals also possess highly specialized, comb-like postcanine teeth, which are used to forage the endemic freshwater amphipod *Macrohectopus branickii* at high rates [15]. Nevertheless, seal hunting continues to this day, mainly targeting molted pups, as the fur of young individuals is considered to be commercially more valuable.

Traditional methods of population size estimation for the Baikal seal remain challenging, due to their cautious nature and a predominantly underwater lifestyle, where they spend 80–90% of their time [5,16–18]. According to different sources, the census population size was estimated at 108,200 individuals in 2013, according to the Russian Ministry of Natural Resources [19], 94,600–137,400 in 2018, and 164,500 in 2021, according to the Baikal branch of the Russian Federal Research Institute of Fisheries and Oceanography (“VNIRO”), “BaikalNIRO” [20]. Estimates based on breeding snow dens are also complicated, as dens are well fortified and camouflaged. Dens have blowholes opening into the water, that are used by females to feed their pups, and are also used by both to escape, in case of danger [7,8]. Seals can be seen above the water surface in the spring, when they use the ice floes to complete their molt and migrate north across the lake. They sometimes lay on rocks near the shore, but only in the spring and summer. However, such individuals usually include only a small percentage of the population (at different times 0.1–10%), often representing tired or sick individuals or those that were not able to complete molt, due to an early ice break [21]. There are many locations suitable for shore haulouts. However, due to negative anthropogenic impact, most of those locations are avoided by seals, which should be regarded as a loss of potential habitat [21].

Climatic changes, and consequent habitat loss or increase, can affect closely related species, especially if they have overlapping areas or occupy similar ecological niches. Baikal seals are now isolated in the lake, but their ancestors were marine animals [3]. Therefore, it makes sense to compare both the modern population size and demographic history of the Baikal seal with other species from the *Phoca-Pusa-Halichoerus* lineage. Currently, whole genome data is available for at least the gray seal (*Halichoerus grypus*), spotted seal (*Phoca largha*), and harbor seal (*Phoca vitulina*). Populations of these species mostly inhabit northern seas and ocean regions (Figure 1). The western Atlantic gray seals are believed to have split from the rest of the population of gray seals at ~0.03 Mya [22]. An expansion of their populations, caused by habitat increase, is believed to have occurred after the last glacial maximum (LGM), at ~0.02 Mya [22]. But in the late 1800s, the eastern Canadian population included only a few dozen individuals, due to harvesting for oil [23]. Nowadays, the numbers have increased to around 450,000 individuals [24]. Since the western Atlantic population went through periods of severe downsizing, and we can exclude trans-Atlantic movement, the modern population must have originated from small remaining populations, and recolonized the coasts of the USA and Canada [25]. The information on spotted seals is rather poor. They are well known to have been harvested for centuries in Russian and Chinese waters, which had a great impact on population size [26]. Since the 1980s, the Bering population has grown from ~248,000 individuals to ~460,000 [27]. However, the southern population is significantly lower, including ~3300 individuals [26]. An interesting feature of gray seals and spotted seals, is their ability to cover huge distances in seasonal feeding, which could lead to genetic exchange between breeding populations and colonies [28–30].

Only a few genetic, and no genomic, studies on Baikal seals have been published. Most of them are based on mtDNA only [2–4,31,32]. However, mtDNA is very short and provides a relatively small number of informative markers. The current trend is to use whole genome data, and chromosome-length assemblies, even in studies of nonmodel organisms [33–36]. Our study fills this gap in the genomics of pinnipeds. We generated the first chromosome-length assembly of the Baikal seal genome. This allowed us to elucidate the genetic diversity, demographic history, and phylogenomics of the Baikal seal, and to compare its features with the additional pinniped species mentioned above.

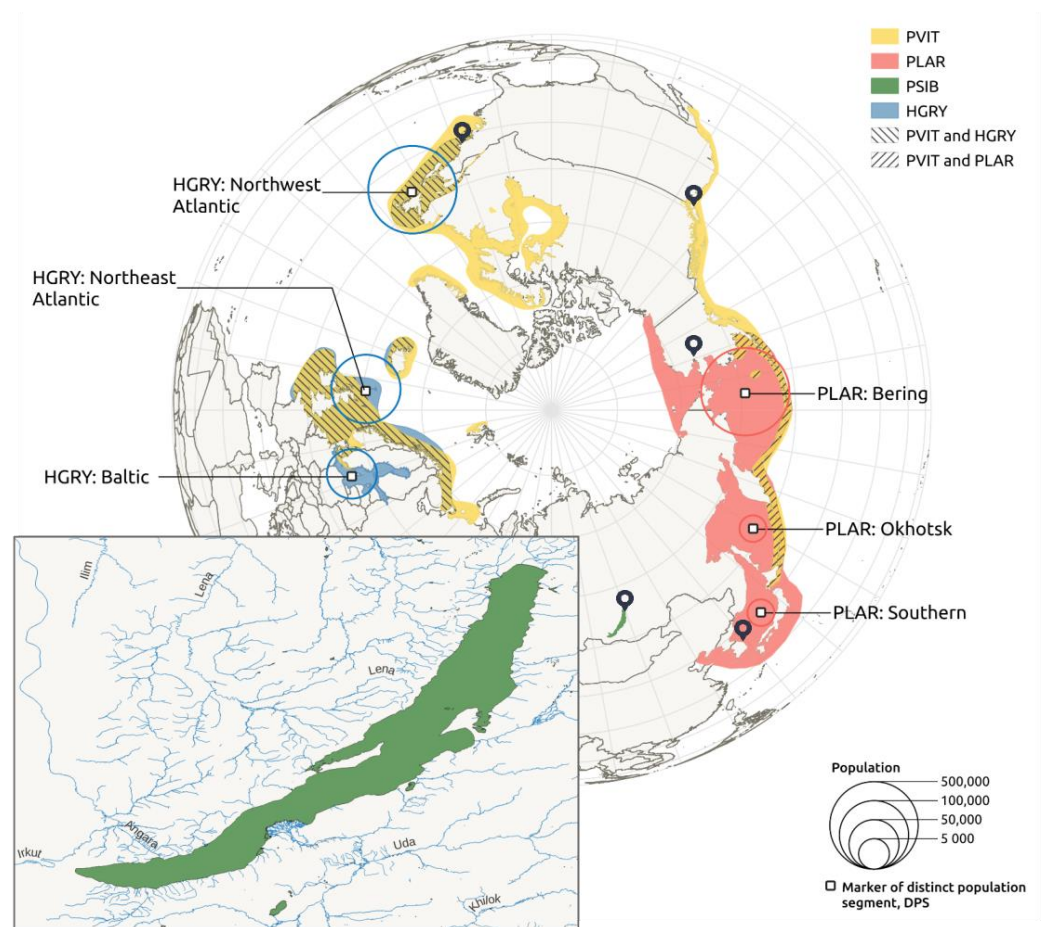


Figure 1. The geographic distributions of the four pinniped species included in this study: Baikal seal (PSIB), gray seal (HGRY), spotted seal (PLAR), and harbor seal (PVIT). Regions of habitat intersection are shown with diagonally shaded areas. Map pointers denote sample coordinates.

2. Materials and Methods

2.1. Sequencing of Baikal Seal Individuals

For whole genome sequencing, we used primary fibroblast cell lines of one male (PUSI1m) and one female (PUSI1f) Baikal seal, obtained from the Novosibirsk Cell Line Collection, located at the Institute of Molecular and Cell Biology, Siberian Branch, Russian Academy of Sciences. Sample collection, transportation, and cell line establishment were previously described in detail in [37].

The male individual was chosen to generate a de novo Baikal seal genome assembly. For this sample, we created one paired-end, and multiple mate pair (jumping), libraries, of different insert sizes, and sequenced these on an Illumina HiSeq 2000 or 2500 (Illumina, Inc., San Diego, CA, USA) instrument. We also generated and sequenced one Hi-C library on an Illumina NovaSeq6000 (Illumina, Inc., San Diego, CA, USA) instrument, which was used to scaffold the assembly to chromosome length. A detailed description of the libraries is provided in Table S1. For the female individual, we generated and sequenced only one paired-end library.

2.2. Filtration, QC, and Postprocessing of Sequencing Data

Quality control of raw and filtered reads was performed using FastQC v0.11.9 [38]. Distributions of 23-mers before and after read filtration were counted using Jellyfish v2.2.10 [39], with parameters ‘-m 23 -s 30G -t 24’ for *jellyfish count*, and ‘-t 4 -l 1 -h 1,000,000 -i 1’ for *jellyfish histo*, and then visualized using KrATER v2.5 (<https://github.com/mahajrod/krater> (accessed on 21 December 2022)), with parameters ‘-m 23 -w 8 -g 150 -u 1’ to check for

possible anomalies and signs of contamination. Trimming of Illumina adapters and quality filtering were performed using Trimmomatic v0.39 [40], with parameters 'ILLUMINA-CLIP:\$adapter:2:30:10 SLIDINGWINDOW:4:15 MINLEN:5'. Baikal seal mate pair libraries were additionally processed using NextClip [41], with default parameters, to remove the Nextera adapter and to extract true mate pair reads. To achieve equal coverage and maximize similarity among samples of all seal species, we cut all reads to 100 bp (the read length used to sequence the Baikal seal samples), and downsampled them to 20× coverage. For downsampling we used FaCut (<https://github.com/mahajrod/Facut> (accessed on 21 December 2022)).

2.3. Assembly of Baikal Seal Reference Genome

A chromosome-length genome assembly of the Baikal seal was generated in three stages. First, we generated a draft assembly from filtered paired-end and mate pair libraries of the reference individual, with the Plantanus v1 genome assembler, using default parameters [42]. Next, we scaffolded the draft assembly to chromosome level, using the reads from the Hi-C library and the 3D-DNA pipeline, with default parameters [43]. Finally, the generated scaffolds were manually verified and corrected using Juicebox [44].

2.4. Genomes of other Pinnipeds and QC of All Assemblies

For comparative analysis, we used chromosome-length assemblies and sequencing data of three related pinniped species [45–48]: the gray seal (SRX12373285), spotted seal (SRX9606535, SRX3524687), and harbor seal. Assemblies and reads were downloaded from the DNA Zoo (<https://www.dnazoo.org/> (accessed on 21 December 2022)) and NCBI Sequence Read Archive (SRA) (Table 1). Genome assemblies of all species were assessed for general quality metrics and gene completeness using BUSCO v5 and the mammalia_odb10 and carnivora_odb10 databases [49].

Table 1. General description and quality metrics for genome assemblies of four pinniped species used in this study. MP = mate pair library, PE = paired-end library.

	Baikal Seal (PSIB)	Gray Seal (HGRY)	Spotted Seal (PLAR)	Harbor Seal (PVIT)
IUCN Red List Category ¹	LC	LC	LC	LC
Gender	male	male	female	female
Assembly source	DNAzoo	DNAzoo	DNAzoo	DNAzoo
Assembly accession/reference	-	Halichoerus grypus_HiC	Phoca largha_HiC	GSC_HSeal_1.0_HiC
Sequencing/assembly approach	MP + HiC	Overlapping PE + HiC	Overlapping PE + HiC	Linked reads + HiC
2n	32	32	32	32
C-scaffolds	16	16	16	16
Scaffold N50, Mbp	151.5	141.6	142.1	152
Number of scaffolds	146,408	388,387	249,399	5311
Total genome length, Gbp	2.35	2.4	2.36	2.36
C-scaffolds length, Gbp	2.29	2.16	2.16	2.32

¹ IUCN Red List category: LC—least concern.

2.5. Whole Genome Alignment and Connection between Assembly and Karyotype

First, we softmasked tandem and interspersed repeats in the genomes of the four pinniped species and the stone marten genome (*Martes foina*), using Tandem Repeats Finder v4.09.1 [50], WindowMasker 1.0.0 [51], and RepeatMasker v4.1.2.p1 [52]. Second, a whole genome alignment of the five analyzed species was performed, using Progressive Cactus [53], with default parameters. Next, synteny blocks were extracted from the multiple whole genome alignment using halSynteny v2.2 [54], with the options—minBlockSize 50000—maxAnchorDistance 50000. Finally, we visualized synteny blocks using ChromoDoter (<https://github.com/mahajrod/ChromoDoter> (accessed on 21 December 2022)) and the draw_synteny.py script from the MACE package (<https://github.com/mahajrod/mace>

(accessed on 21 December 2022)). Dot plots, generated by ChromoDoter, were manually compared to Zoo-FISH experiments previously published for PSIB, HGRY, and PLAR karyotypes, with MFO chromosome libraries used as probes [37,55–58].

2.6. Read Alignment, Coverage Estimation, and Variant Calling

Filtered paired-end reads of analyzed individuals were aligned to the corresponding reference genome assemblies with BWA v0.7.17, using the MEM algorithm [59], followed by duplicate marking, sorting, and indexing using Samtools v2.30.0 [60]. For all generated alignments we calculated per-base coverage using Mosdepth v0.3.1 [61], and visualized it on heatmaps using the MACE package (<https://github.com/mahajrod/MACE> (accessed on 21 December 2022)).

To set the correct ploidy for the X chromosome during variant calling, we identified the pseudoautosomal region (PAR). First, median coverage was determined in stacking windows of 10 kbp. Adjacent windows with median coverage $\geq 70\%$ of the whole genome level were merged. Then all the combined windows with the median coverage of the intermediate gap $\geq 70\%$ of the whole genome level were merged. This procedure improves the accuracy of detecting the PAR coordinates.

SNPs and short indels were detected using bcftools v1.15 [60]. First, we used the *bcftools mpileup* command, with default parameters. Next, the *bcftools call* command was run, with the set PAR coordinates. Finally, low-quality variants were removed with the *bcftools filter* command, with the following parameters: “QUAL < 20.0 | (FORMAT/SP > 60.0 | FORMAT/DP < 5.0 | FORMAT/GQ < 20.0)”. Finally, only variants with coverage in the 30–250% range, relative to the mean whole genome coverage, were retained.

2.7. Heterozygosity

For analysis of heterozygosity, we applied a sliding window approach. Heterozygous SNPs were counted in overlapping sliding windows of 1 Mbp, with a step size of 100 kbp, and counts were scaled to SNPs per kbp. Heterozygosity counts were visualized on chromosome scaffolds and violin plots, using the MACE package and Matplotlib library (<https://matplotlib.org> (accessed on 21 December 2022)).

2.8. Demographic Reconstruction

Demographic history inference was performed using PSMC v0.6.5 [62], with the following parameters: -N25 -t15 -r5 -b -p '4 + 25 × 2 + 4+6'. We used mutation rate and generation time values based on scientific publications of the target species, or from closely related species [22,63–67]. For mutation rate, we used values of 0.7×10^{-8} , 1.2×10^{-8} , and 2.5×10^{-8} substitutions per site per generation, which is in the range of commonly used values for mammals [68–70]. Generation times were chosen to be equal to 21.6 years for the Baikal seal [19], 16.5 years for gray seal [71], and for spotted seals we used 14.8 years, which is the same as commonly used for its close relative, the harbor seal [72,73]. Effective population size trajectories for each species were visualized with the *psmc_plot* command.

2.9. Phylogenetic Tree Reconstruction and Dating

We reconstructed a phylogenetic tree using multiple alignments of coding sequences of single-copy orthologous genes (BUSCOs). In the reconstruction, we included 11 species: cheetah (*Acinonyx jubatus*), mountain lion (*Puma concolor*), domestic dog (*Canis lupus familiaris*), brown bear (*Ursus arctos*), red panda (*Ailurus fulgens*), walrus (*Odobenus rosmarus*), bearded seal (*Erignathus barbatus*), harbor seal (*Phoca vitulina*), spotted seal (*Phoca largha*), gray seal (*Halichoerus grypus*), and Baikal seal (*Pusa sibirica*). Corresponding assemblies were downloaded from the NCBI Genome database (<https://www.ncbi.nlm.nih.gov/genome/> (accessed on 21 December 2022)) and DNA Zoo (<https://www.dnazoo.org/> (accessed on 21 December 2022)): AJUB, ID:aciJub1_HiC [74]; PCON, ID:PumCon1.0_HiC [75]; CLUP, ID:canFamDis_HiC [76]; UARC, ID:ASM358476v1_HiC [77]; AFUL, ID:ASM200746v1_HiC [78]; OROS, ID:Oros_1.0_HiC [79]; EBAR,

ID:Erignathus_barbatus_HiC [80]; PVIT, ID:GSC_HSeal_1.0_HiC [81]; PLAR, ID:Phoca_largha_HiC [80]; HGRY, ID:Halichoerus_grypus_HiC [43,44,80]. BUSCO sequences were generated using BUSCO v.5.4.2 [49] with the Mammalia_odb v.10 (2021-02-19) database of orthologs (9226 BUSCOs). Only single-copy sequences common for all species were included in the analysis. Multiple codon alignments were performed separately for each ortholog using PRANK v.170427 [82], followed by filtration of the hypervariable and poorly aligned regions using GBlocks v.0.91b [83]. A phylogenetic tree, based on the concatenated alignments of 5025 BUSCOs (8,153,886 Mbp), was estimated using the maximum likelihood method implemented in the IQ-Tree v.2.2.0 [84], with automatic selection of the best-fitting DNA substitution model using ModelFinder [85]. For the evaluation of node support, 1000 bootstrap replicates were generated.

Dating of the phylogenetic tree was performed using the MCMCtree tool from the PAML package v4.7 [86], with the HKY85+G model of nucleotide substitution and 220,000 MCMC generations, of which the first 20,000 generations were discarded as burn-in. Divergence times were assessed using 4-fold degenerate sites, which were extracted from the concatenated codon alignment generated during the tree reconstruction stage. Fossil calibrations used for dating are listed in Supplementary Table S7. The final dated tree was visualized using FigTree v.1.4.4 (<http://tree.bio.ed.ac.uk/software/figtree> (accessed on 21 December 2022)).

3. Results

3.1. Genome Size and Assembly, Repeat Content

We assembled the first chromosome-length reference genome of a male Baikal seal (PSIB) individual using 2386.3 million paired-end reads, 735.1 million mate pair reads of different insert sizes, and 1348.5 million Hi-C reads (Table S1). An estimation based on the distribution of 23-mers extracted from filtered paired-end reads, showed a genome size of 2.12 Gbp for the male individual, and 2.22 Gbp for the female (Supplementary Figure S1A). Downsampling to the standardized 20× coverage did not change the estimates (Supplementary Figure S1B). Two closely related species, the gray seal and spotted seal, for which raw sequencing data were available, demonstrated similar values: 2.29 Gbp and 2.37 Gbp, respectively. The Baikal seal assembly had a total length of 2.35 Gbp, which is slightly higher than the genome size estimate. Scaffold N50 reached 151.5 Mbp (Table 1), and we observed a dramatic 29-fold difference in length between the 16th (57.63 Mbp) and 17th (1.99 Mbp) longest scaffolds. Thus, our assembly includes 16 chromosomal scaffolds (C-scaffolds), which exactly fit the number of chromosome pairs in the karyotype ($2n = 32$). A BUSCO-based assessment indicated high completeness of the assembly. For the mammalia_odb10 (9226 BUSCOs) and carnivora_odb10 (14,502 BUSCOs) databases, we found 86.5% and 88.0% complete and single-copy, and 1.5% and 1.6% complete and duplicated BUSCOs, respectively. These, and other metrics for previously published pinniped species, are somewhat higher, yet comparable (Table 2, Supplementary Table S3). Interspersed repeat content among pinniped species is also similar, with LINEs, SINEs, and LTRs encompassing 3.16%, 19.18%, and 4.92%, respectively, of the Baikal seal genome (Table S4).

Table 2. Completeness of pinniped genome assemblies assessed using BUSCO v5; the mammalia_odb10 database of 9226 orthologs was used in the analysis.

Species/BUSCOs	Complete Single-Copy	Complete Duplicated	Fragmented	Missing
<i>P. sibirica</i>	7981 (86.5%)	140 (1.5%)	444 (4.8%)	661 (7.2%)
<i>H. grypus</i>	8352 (90.5%)	190 (2.1%)	242 (2.6%)	442 (4.8%)
<i>Ph. largha</i>	8434 (91.4%)	178 (1.9%)	215 (2.3%)	399 (4.4%)
<i>Ph. vitulina</i>	8651 (93.8%)	198 (2.1%)	90 (1.0%)	287 (3.1%)

3.2. Connection between Assembly and Karyotype

Of the four analyzed species, Zoo-FISH data is available only for the Baikal seal and harbor seal, making it possible to assign C-scaffolds to particular chromosomes in the karyotype for these two species. Following identification, C-scaffolds were assigned to the corresponding chromosome numbers with the prefix “chr”. For the two other species (HGRY and PLAR), such an assignment was possible only for the X chromosome, because it is highly conserved across Carnivora and mammals [87]. The remaining C-scaffolds were numbered from longest to shortest, with the prefix “aut” indicating “autosomal” (Supplementary Table S5). For the Baikal seal and spotted seal, we found no contradictions between the Zoo-FISH data and whole genome alignments (WGA).

Synteny blocks (>50 kbp in length) extracted from the whole genome alignment, showed a high similarity between PSIB, HGRY, PLAR, and PVIT (Figure 2). Species with assigned chromosomes (Baikal seal and harbor seal) have precise correspondence between chromosome ids, i.e., chr1 of one species corresponds exactly to chr1 of the second. However, C-scaffolds of several chromosomes (chr2, chr4, chr3, chr7, chr10, chr13, chr14, chr15, and chrX) are inverted in the assemblies. By ordering autosome C-scaffolds by length in the spotted seal (PLAR) and gray seal (HGRY), we found them to be slightly rearranged: aut1 and aut2 correspond to chr2 and chr1 of the other species, and aut13 and aut14 correspond to chr14 and chr13, respectively.

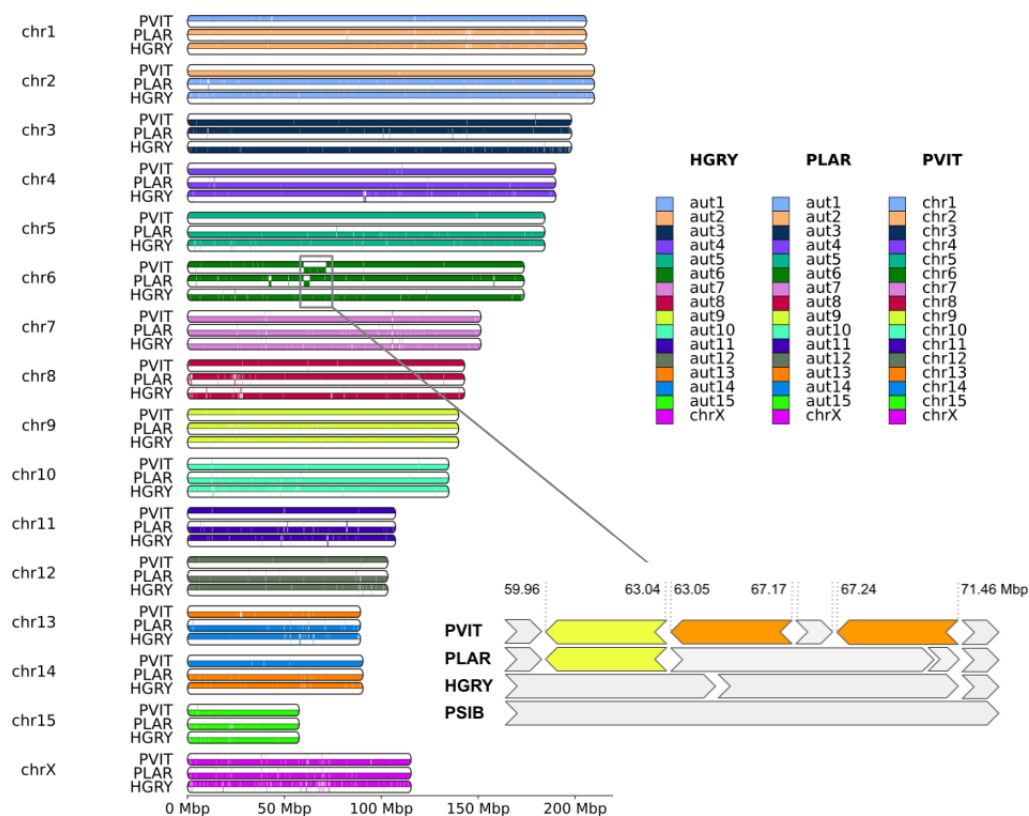


Figure 2. The synteny blocks between four pinniped species: PSIB was chosen as a reference and HGRY, PLAR, and PVIT were designated as the query species. Each chromosome was split into bottom and top parts: synteny blocks were drawn on the top part if it had the same orientation in both the reference (PSIB) and query species, and in the bottom part if the orientation was different. Chr 6 was enlarged, to emphasize the presence of three megabase-scale inversions. The yellow color on the enlarged pane highlights the single inversion common for PVIT and PLAR, and the orange color indicates the two inversions unique to PVIT. HGRY chr6 was reverse complemented in the small pane, and labeled with an asterisk on the small pane to avoid ambiguity.

We detected dozens of short inversions (Supplementary Figure S1) and only three relatively large inversions of megabase-scale on chr 6. One of them (Figure 2, enlarged pane, yellow) is common to the harbor seal (PVIT) and spotted seal (PLAR), and is probably a synapomorphy of the genus *Phoca*, which includes only these species. The second and third inversions (Figure 2, enlarged pane, orange) are unique to the harbor seal (PVIT).

3.3. Heterozygosity and SNP Density

Using a coverage-based approach, we verified the morphological sex of the studied individuals, and detected coordinates of the pseudoautosomal region (PAR) in the male Baikal seal and male gray seal. In both species, the PAR encompasses slightly different lengths at the ends of the X chromosomes: 6.75 Mbp in the Baikal seal and 5.88 Mbp in the gray seal (Figure S2). Heterozygous SNPs, counted in sliding windows of 1 Mbp, with a 100 kbp step size, showed a median heterozygosity (hetSNPs per kbp) of 0.48 for the gray seal, 0.67 and 0.62 for spotted seals, and 0.61 and 0.66 for male and female Baikal seals, respectively (Figure 3F). Mean values did not differ significantly from the median ones (Table 3). Exclusion of the X chromosome does not show any remarkable changes in values: 0.5 for gray seals, 0.68 and 0.63 for spotted seals, and 0.62 and 0.67 for male and female Baikal seals, respectively (Supplementary Table S6).

Table 3. Statistical metrics of heterozygosity for analyzed samples of three pinniped species. SNPs are counted in 1 Mbp sliding windows, with a 100 kbp step size, and scaled to SNPs per kbp.

Species	Min	Median	Mean	Mode	Max
<i>H. grypus</i> , male	0	0.48	0.49	0.48 ¹	1.72
<i>Ph. largha</i> , female (DNAZoo)	0	0.67	0.68	0.66	3.23
<i>Ph. largha</i> , female (Seoul University)	0	0.62	0.62	0.60	2
<i>P. sibirica</i> , male	0	0.61	0.63	0.56 ¹	2.17
<i>P. sibirica</i> , female	0	0.66	0.68	0.56	2.34

¹ For male samples we observed an additional mode at 0. It corresponds to the hemizygous region (the part of the X chromosome outside of the PAR) in males.

Heatmaps of SNP density distribution showed a slightly higher number of heterozygous SNPs on the ends of chromosomes in all individuals, and relatively short regions (dark blue) of low heterozygosity in the middle of several chromosomes (Figure 3). The PAR region had SNP densities comparable to the autosomes in both males and females. Overall, however, SNP densities were generally similar among the three seal species we compared.

3.4. Demographic Reconstruction

A PSMC-based reconstruction of the historical demography for the Baikal seal, gray seal, and spotted seal resulted in different trajectories of effective population size (N_e) changes, but individuals from the same species showed very similar tracks (Figure 4). The gray seal, spotted seal, and Baikal seal share one significant N_e peak in the time intervals ~1–0.8/1.5–2/3–4 Mya (here and below we provide dating for three mutation rates in the following order: 2.5×10^{-8} / 1.2×10^{-8} / 0.7×10^{-8}). They also share a second peak that is more extended over time, and varies in intensity among the three species: for the gray seal it is higher, while for the spotted seal it is lower, with both occurring at ~0.2/0.4/0.7 Mya with respect to μ . At the bottom of the first population decline, ~0.3/0.6/1 Mya, N_e decreased by ~40% in the gray seal and ~75% in the spotted seal. For Baikal seals, at ~0.8/1.8/3 Mya, the most dramatic decline had started, with a roughly 10-fold decrease in N_e , according to the demographic reconstruction. After the second, relatively small decrease, the spotted seal population stabilized at ~0.1/0.2/0.4 Mya, and even showed a trend for growth, but the N_e for the gray seal continued to decrease until modern times. The decline in population size for the Baikal seal was followed by a small growth in N_e at ~0.25/0.5/0.9 Mya, followed by stabilization at that level.

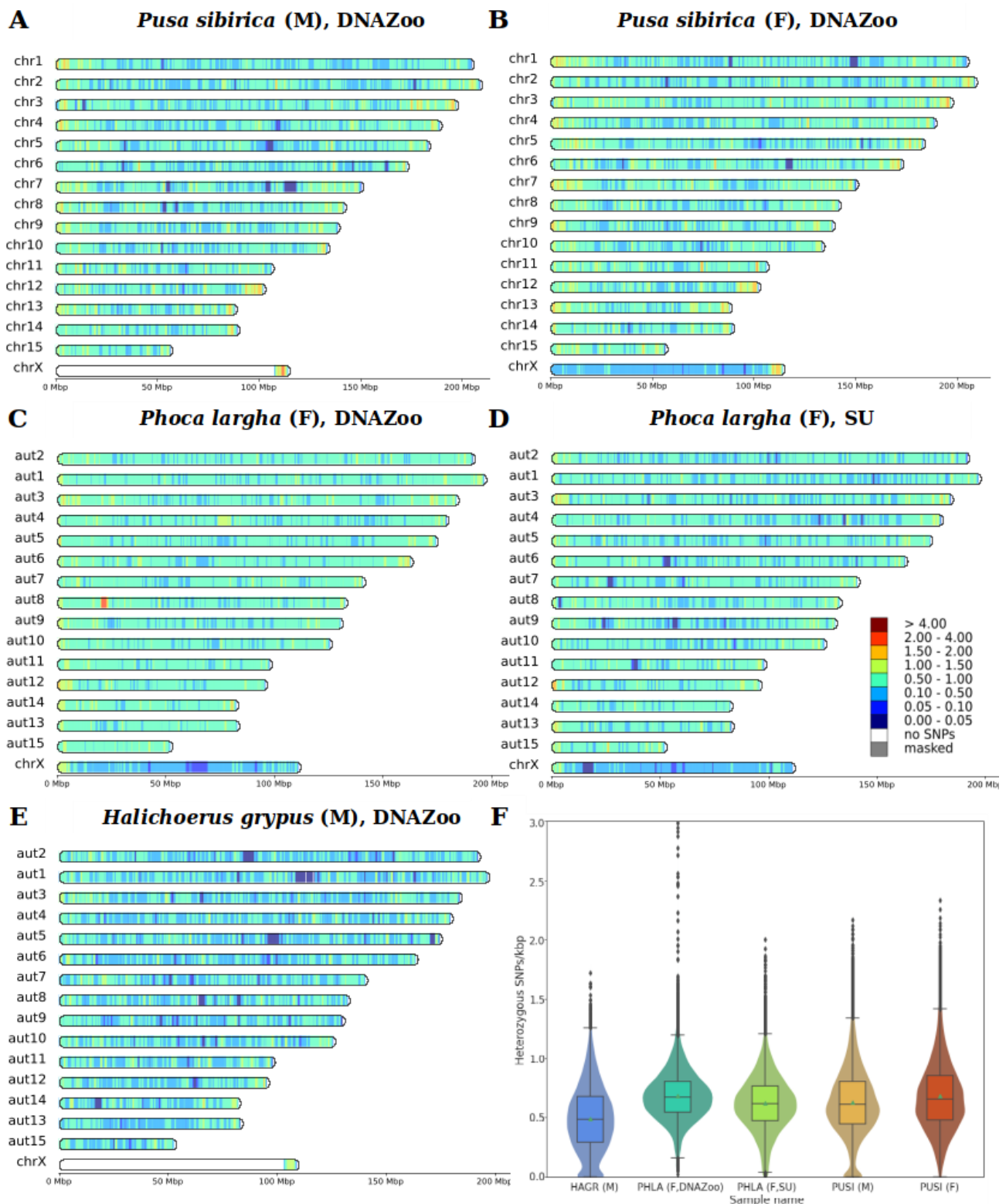


Figure 3. Heterozygous SNP densities (SNPs/kbp) for analyzed samples of three pinniped species. HetSNPs were counted in 1 Mbp sliding windows, with a 100 kbp step size, and scaled to SNPs/kbp. (A) PSIB, male, (B) PSIB, female, (C) HGRY, male, DNA Zoo, (D) PLAR, female, DNA Zoo, (E) PLAR, female, Seoul University, (F) violin and boxplots of heterozygosity distribution for all five pinniped samples. Autosomes of PLAR and HGRY were arranged according to their correspondence to PSIB and PVIT chromosomes.

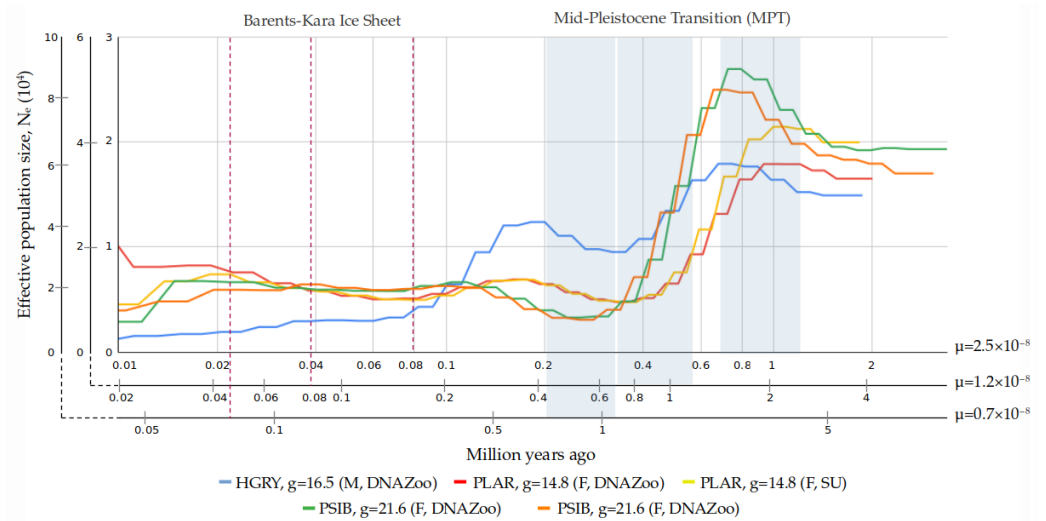


Figure 4. Demographic history reconstruction for all available samples of PSIB, HGRY, and PLAR. Three time scales were used for each value of mutation rate ($\mu = 2.5 \times 10^{-8}$ substitutions per site, $\mu = 1.2 \times 10^{-8}$, and $\mu = 0.7 \times 10^{-8}$). Generation time (years) was set at different times between the species: PSIB—21.6 years, HGRY—16.5 years, PLAR—14.8 years.

3.5. Phylogenomics and Molecular Dating

We reconstructed and dated a maximum likelihood phylogenetic tree of five phocid seals and six other carnivore species, using five fossil calibrations (Supplementary Table S7). Two felid species (cheetah and jaguarundi) were used as the outgroup to root the tree and to provide a fossil-dated root. All nodes on the tree were supported at 100% based on 1000 bootstrap replicates. According to our results, HGRY and PSIB are sister species, whereas *Phoca* (PLAR, and PVIT) form an outside group (Figure 5). The divergence between HGRY and PSIB was assessed to occur around ~1.6 Mya, with a 95% confidence interval (CI) encompassing 2.2–1.1 Mya. For PLAR and PVIT, speciation occurred later, at ~0.7 Mya (95% CI 1–0.4 Mya). The division between *Phoca* and *Halichoerus-Pusa* lineages was dated to ~2 Mya, with a 95% CI of 2.6–1.4 Mya.

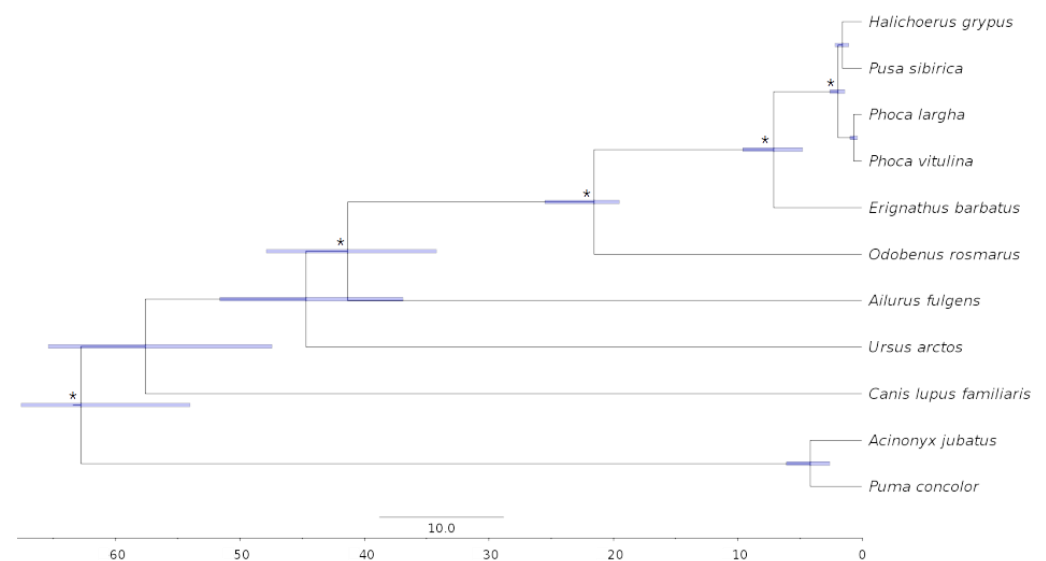


Figure 5. Dated phylogenetic tree of 11 carnivore species, including five phocid seals (HGRY, PSIB, PLAR, PVIT, and EBAR). Blue bars show the 95% confidence intervals. Asterisks (*) label nodes with fossil-based calibration priors (Supplementary Table S8). All nodes were supported at 100% based on 1000 bootstrap replicates.

4. Discussion

4.1. Chromosome-Level Assemblies and Karyotypes

Assignment of chromosome numbers to C-scaffolds can only be established if Zoo-FISH or other marker-based mapping data is available [88]. Such data were previously published for the Baikal seal and harbor seal. Our findings show that chr1 in the assemblies (specifically, the C-scaffold) of both species is shorter than chr2. However, in the karyotypes, chr1 is longer than chr2. Such a contradiction may be explained by the presence of an unassembled heterochromatin region in the short arm of chr1, between segments homologous to parts of dog chr5 and chr9 [37]. This region is clearly visible using both C- and G-banding methods, and was identified to be associated with the nucleolus organizer region (NOR) [56]. The NOR consists of multiple ribosomal DNA (rDNA) copies and is impossible to assemble at full length from short reads. Usually, only a few copies of rDNA monomer fragments are present in genome assemblies. Manipulations of reads and contigs, and even additional sequencing (preferably using long read sequencing), are required to generate the correct sequence of the rDNA monomer [89].

Other chromosomes possibly not ordered by length, include chr13 and chr14. The length of these chromosomes seems to be similar, both in the assemblies and karyotypes. The most likely explanation for the discordance is the limited precision for estimation of chromosome length. For the gray seal and spotted seal, we lack Zoo-FISH data. To emphasize this difference, and issues with chr1, chr2, chr13, and chr14, we used the prefix “aut” (instead of “chr”) for C-scaffolds of the two latter species, and assigned numbers to them according to their length (Figures 2 and 3), using the standard rules of chromosome nomenclature [90,91].

4.2. Genome Rearrangements and Speciation

The genus *Phoca* contains only two species, the harbor seal (PVIT) and spotted seal (PLAR), both of which were included in our analysis. We detected a single four megabase synapomorphic inversion for this genus, and two more inversions of the same scale specific for the harbor seal. All three inversions are located nearby to one another, on chromosome 6. These findings immediately raise the question of whether these genome rearrangements were associated with divergence of the genus *Phoca*, and speciation within it. Inversions are well known to partially suppress crossover events [92] within themselves, due to unsustainable meiotic products. This leads to linkage disequilibrium [93] and reduced fertility of individuals with heterozygous inversions, and might lead to partial reproductive isolation [94–96]. Multiple or nested inversions may aggravate the effect [97]. The second result of such rearrangements, is a significant change in the genome environment, which might affect expression [98], with multiple possible consequences in phenotype or behavior, depending on what genes are included in the inversion. The spotted seal, harbor seal, and gray seal have partly overlapping ranges (Figure 1), and reproductive isolation based on genetic background might shape the speciation within the Phocidae [99]. On the other hand, the detected inversions might be just a genetic feature of the specific individuals that were sequenced. This is less likely for the *Phoca*-specific inversion, as it was detected in two different species, but for the two other inversions detected in the harbor seal, additional individuals representing different populations should be sequenced to further explore the frequency of these inversions.

4.3. Heterozygosity

Heterozygosity in pinniped species is known to be low [22,26,100]. According to our results, the Baikal seal, the only pinniped among those analyzed that occurs within a landlocked habitat, showed a mean heterozygosity of 0.63 and 0.68 SNP/kbp for male and female individuals, respectively, comparable to that found in marine pinniped species. Moreover, it was even higher than for gray seals (0.49 SNP/kbp) (Figure 2A,B,E). Low heterozygosity is often associated with reduced fertility and survival rate, and might be a forerunner of further decline, or even extinction [101]. However, nowadays the population

trend of Baikal seals is recognized as stable, and the species has the status of “least concern” (LC) on the IUCN Red List of Threatened Species [19]. By comparing heterozygosity levels of representative species from the *Felidae*, *Ailuridae*, *Mustelidae*, and *Phocidae*, we observe that pinnipeds are the only species to have a low heterozygosity level without being listed within a threatened status category (Figure 6).

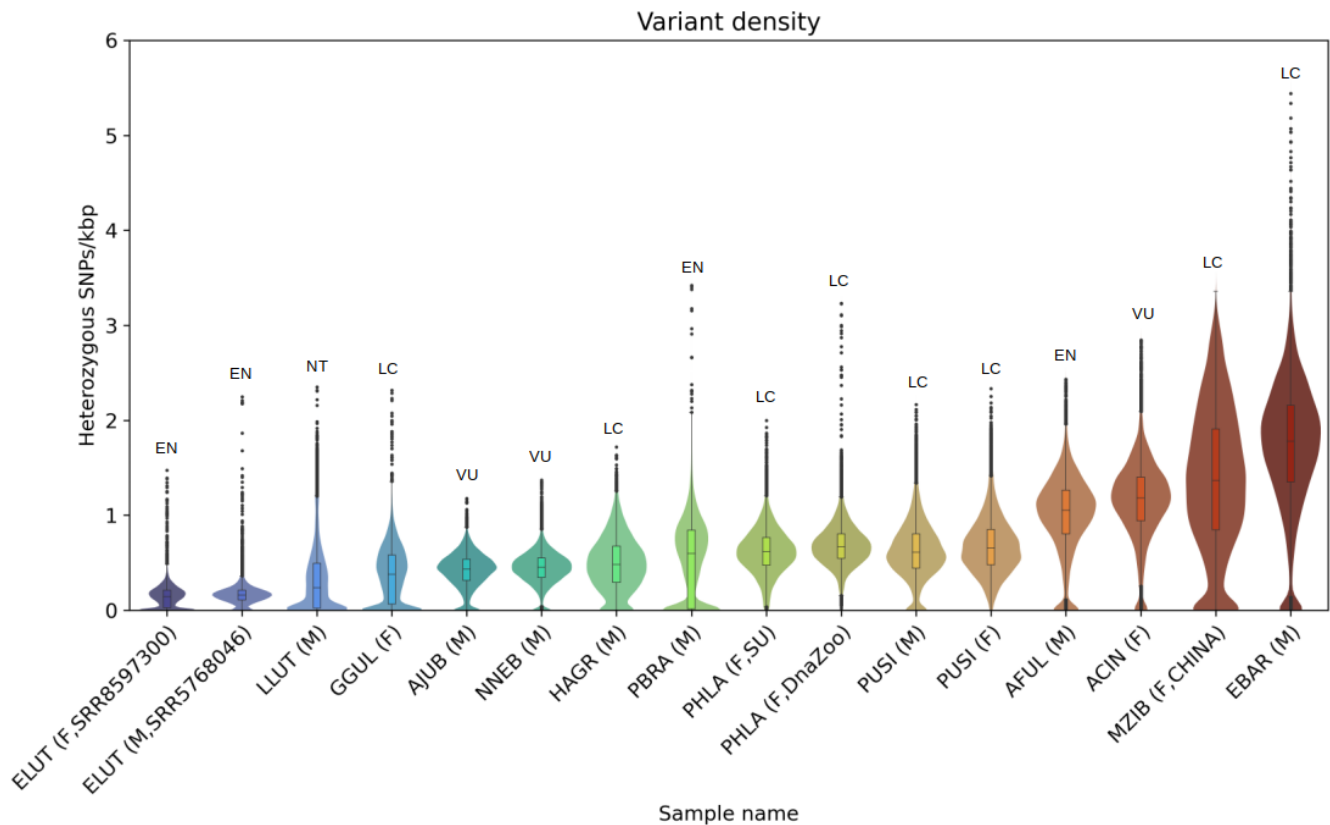


Figure 6. Violin and boxplots of heterozygosity distribution for analyzed samples of *Felidae*, *Ailuridae*, *Mustelidae*, and *Phocidae* species: *Enhydra lutris* (ELUT, EN), *Lutra lutra* (LLUT, NT), *Gulo gulo* (GGUL, LC), *Acinonyx jubatus* (AJUB, VU), *Neofelis nebulosa* (NNEB, VU), *Halichoerus grypus* (HAGR, LC), *Pteronura brasiliensis* (PBRA, EN), *Phoca largha* (PHLA, LC), *Pusa sibirica* (PUSI, LC), *Ailurus fulgens* (AFUL, EN), *Aonyx cinereus* (ACIN, VU), *Martes zibellina* (MZIB, LC), *Erignathus barbatus* (EBAR, LC). IUCN Red List categories: LC—least concern, NT—near threatened, VU—vulnerable, EN—endangered.

Reduced heterozygosity can be a sign of extensive inbreeding in an enclosed ecosystem. Baikal seals are effectively trapped in the lake, but Lake Baikal occupies a vast area (with a surface area of 31,722 km²), with a very diverse and endemic flora and fauna. Current census count estimates vary between 82,500 and 115,000 seals, which demonstrates the sufficient capacity and resources of the lake to support a large population of a large-bodied predator such as the Baikal seal [15,19]. For comparison, the population of ringed seals, whose area encompasses the entire Arctic Ocean and northern parts of the Atlantic and Pacific Oceans, was estimated to reach “only” ~1.5 million individuals [73]. Another possible explanation to consider for the reduced heterozygosity, is that Baikal seals could have experienced a decline in their initial population size, and hence their genetic diversity, during their long migration to, and isolation in, Lake Baikal. The relatively uniform distribution of heterozygous sites across the genome (Figure 2A,B) may indicate that the current population descended from only a small group of animals, that separated from their ancestors and settled the current habitat [100].

4.4. Interpretation of Demographic History Reconstruction

The timescale for PSMC-based reconstruction of demographic history is dependent on the ratio of two parameters: generation time and mutation rate (g/μ). Neither parameter is known with certainty for the Baikal seal. We used a mutation rate (μ) of 2.5×10^{-8} substitutions per site, based on the available data for cetaceans and what is commonly used for mammals, 1.2×10^{-8} for medium-sized mammals with approximately the same body size as seals, and 0.7×10^{-8} for pinnipeds [69,102]. Smaller mutation rates result in stretching out the timeline and causing changes in N_e to be dated older (Supplementary Figure S4). For generation times, we used values from the IUCN Red List of Threatened Species, which are available for all our species except spotted seals. Under this standard, generation length is defined as “the average age of parents of the current cohort, reflecting the turnover rate of breeding individuals in a population” [103].

The strong connection of Baikal seals to ice suggests an Arctic origin of the species [2,104]. According to multilocus and mitogenome-only phylogenies [3,31], the closest relative of the Baikal seal is the ringed seal, which inhabits the Arctic Ocean and several sub-Arctic seas, such as the Baltic Sea. Ringed seals have a similar association with ice, the same structure of snow dens, and can maintain breathing holes through at least 1.8 m (6 feet) of ice [105]. Such facts make it reasonable to speculate about what part of the Baikal seal’s reconstructed demographic trajectory is shared with that of the ringed seal via recent common ancestry. These two species are estimated to have diverged ~ 1.15 Mya (95% CI: 1.6–0.72 Mya) [31] or ~ 1.5 – 1.7 Mya [3]. However, such a hypothesis could be verified only after whole genome sequencing of ringed seal individuals. But glacial–interglacial cycles during the Pleistocene likely had an effect on the demographic history of all seals, including the Baikal seal [99].

The Mid-Pleistocene Transition (MPT) lasted from approximately 1.25 Mya to 0.7 Mya [106,107]. Ocean sediments show that during the MPT, there occurred a shift in terminations, switches from glacial to interglacial (G–IG) climates, from a symmetric 41 ky to an asymmetric 100 ky periodicity [106,107]. An increase in the amplitude of G–IG conditions was also characteristic for the post-MPT interval, which consequently led to thickening of ice sheets during glaciation periods and an increase in sea-surface and Antarctic interglacial temperature [108,109]. The demographic reconstruction performed with all chosen values of μ could explain the dramatic downturn of effective population size in the different seal species during that period (Figure 4, Supplementary Figure S4). However, the broad range of the available μ assessments makes the association between bottlenecks and specific glaciation events ambiguous.

For the Baikal seal, the maximum decline encompassed up to 80% of population size, and is the most severe bottleneck observed among the studied seal species. It might indicate the differences in habitat among the seal species at that time. The advancing ice line may have forced the migration of ancestors of Baikal seals and driven them into rivers. During the Early Pliocene, the sea ice appeared at ~ 4.5 Mya at the level of Iceland [110], it became widespread during Northern Hemisphere glaciation at ~ 2.75 Mya [111], and after 1.5 Mya the ice sheet expanded repeatedly towards the continental shelf [112] along the western Barents Sea and Svalbard margins [113–115]. Based on $\mu = 1.2 \times 10^{-8}$ and $\mu = 0.7 \times 10^{-8}$, we could suppose that the migration took place between ~ 1.5 – 0.6 and ~ 3 – 1 Mya, respectively. This period corresponds to the time of ice expanding and conform the earlier studies [2,3]. According to the seismic structure of the northwestern margin, at least sixteen glacial advances occurred during the last 1 Myr [116]. In addition, a path between the Baltic Sea and White Sea, through Lake Ladoga, appeared and disappeared multiple times during this period [117,118]. At least, during the Eemian interglacial period (115–130 ka ago), all this area was covered by a single sea basin, desalinated due to river inflow [118,119]. It is an outstanding question of when isolated regions were completely covered or not by ice, but theoretically they could be inhabited by isolated populations of seals. If we take into account $\mu = 2.5 \times 10^{-8}$, after the first massive peak at ~ 0.7 Mya, the decline in effective population size of the Baikal seal was reached between ~ 0.4 – 0.2 Mya.

We could suggest that migration to Lake Baikal started at some time between ~0.7–0.3 Mya. A similar date (~0.4 Mya) for the migration to Lake Baikal was proposed earlier, using an assessment of mtDNA-based genetic distance between *Pusa* species [4]. Another significant event possibly related to the history of the Baikal seal, was when the Yenisey and Ob rivers were blocked by the Barents–Kara Ice Sheet, at approximately 0.08 Mya, and the West Siberian plain was covered by the West Siberian Glacial Lake at this time [120]. Reconstructions for all values of mutation rate showed stabilization of effective population size before ~0.1 Mya. (Figure 4). This might indicate that Baikal seals inhabited the lake since at least that period, and endured the Last Glacial Period (LGP), lasting from 115 ka until 11.7 ka [121–123].

4.5. Phylogenetic Analysis

The phylogeny of the true, or earless, seals (Phocidae) to date is considered to be mostly resolved, based on molecular and morphological data [3,32]. However, there are controversial relationships between some species, despite the abundance of genetic data. In particular, the placement of species among the *Phoca*–*Pusa*–*Halichoerus* lineages is not well resolved [32]. One of the reasons why it is a challenge to resolve the phylogeny of these species, could be due to the rapid radiation of northern ancestors over vast areas, and the formation of large populations with overlapping habitats. The speed of radiation, and thus speciation, could be confirmed by the phylogenetic tree we obtained, where divergence events are tightly spaced in time. According to our analysis, divergence between *Phoca* and *Pusa*–*Halichoerus* happened ~2 Mya (95% CI 2.6–1.4 Mya), and given that generation lengths of selected species are ~15–22 years, one can assume that sufficient effective generations have not passed for complete lineage sorting [32]. That might also be indicated by the high genome similarity of gray and Baikal seals, despite their different habitats, isolation from each other, and morphological differences [32]. Some researchers have even suggested merging all three genera into a single genus, *Phoca* [3]. The estimated divergence times are consistent with the previously estimated divergence within *Pusa*, and occurred within 1.7–0.8 Mya, based on Bayesian dating using 15 nuclear genes and near-complete mitochondrial genome sequences [3,31].

5. Conclusions

This work was to resolve the problem of the migration of the Baikal seal to Lake Baikal using the assembled genome. However, despite providing new insights, our study raises more questions. The important issue is the specific demographic trajectory of the Baikal seal. Potentially, it could be further elucidated through genome assembly followed by demographic reconstruction for the ringed seal (Arctic Ocean population) and Caspian seal. The Ladoga ringed seal and Saimaa ringed seal are less important, as they are known to descend very recently from the Baltic ringed seal [65]. In addition, whole genome sequencing of the entire *Phoca*–*Pusa*–*Halichoerus* lineage will resolve the remaining issues with phylogeny and dating of speciation events. Comparing the gray, harbor, spotted and Baikal seal genomes, we revealed several interesting megabase-scale inversions in chr6 in the *Phoca* lineage, which might be related to speciation. However, they might be just a feature of reference individuals, and only a population scale study will clarify this issue. This can be solved by sequencing 8–10 more samples of each species. Sequencing data of different populations can help to model demographic scenarios and gene flow among populations. At this stage, our assembly will provide a resource for a population genomic study of Baikal seals that will help inform ongoing conservation efforts for this species, and help to preserve its genetic diversity and understand the potential burden of mutational load.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/genes14030619/s1>, Table S1: Description of Baikal seal libraries; Table S2. Read counts of *Pusa sibirica* and additional pinniped samples before and after trimming; Table S3. Genome completeness metrics based on orthologs analysis with BUSCO ver.5, using

carnivora database (carnivora_odb10); Table S4. Major types of transposable elements detected in the genome of Baikal seal; Table S5. Chromosome assignments; Table S6. Statistical metrics of heterozygosity for analyzed samples of three pinniped species, excluding X chromosome. SNPs are counted in 1 Mbp sliding windows with 100 kbp step, and scaled to SNPs per kbp; Table S7. Table with calibrations used for dating in phylogeny analysis; Table S8. Dating of divergence times based on dated maximum likelihood phylogenetic tree of five seals (*Erignathus barbatus*, *Phoca vitulina*, *Phoca largha*, *Halichoerus grypus*, *Pusa sibirica*) and six other carnivore species (*Acinonyx jubatus*, *Puma concolor*, *Canis lupus familiaris*, *Ursus arctos*, *Ailurus fulgens*, *Odobenus rosmarus*) using five fossil calibrations; Figure S1. K-mer distribution of 23-mers for *Pusa sibirica*, *Halichoerus grypus*, and *Phoca largha* read after standard step of adapters trimming (A), and after further cut length, and downsampling (B). PUSI_M—*Pusa sibirica* male sample; PUSI_F—*Pusa sibirica* female sample; HAGR_M—*Halichoerus grypus* DNAZoo sample; PHLA_F_DZ—*Phoca largha* DNAZoo sample; PHLA_F_SU—*Phoca largha* Seoul University sample; Figure S2. Coverage plots for (A) *Pusa sibirica*, female, DNAZoo, (B) *Pusa sibirica*, male, DNAZoo, (C) *Halichoerus grypus*, male, DNAZoo, (D) *Phoca largha*, female, DNAZoo, and (E) *Phoca largha*, female, Seoul University. Coverage was calculated in non-overlapping sliding windows of 1 Mbp and divided by whole genome median coverage; Figure S3. Genome-wide heterozygosity distribution for two samples of Baikal seal (*Pusa sibirica*), one sample of gray seal (*Halichoerus grypus*), and two samples of spotted seal (*Phoca largha*). SNPs are counted in 1 Mbp non-overlapping sliding windows and scaled to heterozygous SNPs per kbp; Figure S4. Demographic history reconstruction for all available samples of *Pusa sibirica*, *Halichoerus grypus*, and *Phoca largha* with mutation rate 2.5×10^{-8} excluding X chromosome; Figure S5. The phylogenetic tree for eleven carnivores, including five seal species (*Halichoerus grypus*, *Pusa sibirica*, *Phoca largha*, *Phoca vitulina*, and *Erignathus barbatus*). Node labels show posterior probabilities. Branch lengths are in proportion to expected changes per site.

Author Contributions: Conceptualization, S.J.O., A.G. and S.K.; methodology, A.Y., P.D., A.K., V.B., K.K., P.L.P., N.C., A.L., DNA Zoo Consortium, K.-P.K. and S.K.; software, A.Y., A.T. (Andrey Tomarovsky), N.C., K.K., G.T., A.K. and S.K.; validation, A.Y., V.B., M.L., A.P., P.L.P., P.D., N.A.S., A.T. (Andrey Tomarovsky), A.T. (Azamat Totikov), E.B., M.R., G.T., K.-P.K. and S.K.; formal analysis, A.Y., M.L., V.B., P.L.P., A.P., A.T. (Andrey Tomarovsky), K.K., N.A.S., M.R., G.T., A.K. DNA Zoo Consortium, and S.K.; investigation, A.Y., V.B., V.P. (Vladimir Pylev), V.P. (Vladimir Peterfeld), M.L., P.L.P., P.D., A.T. (Andrey Tomarovsky), M.L., K.K., A.K., G.T., M.R. and S.K.; resources, V.B., M.L., P.L.P., N.A.S., N.C., V.P. (Vladimir Pylev), V.P. (Vladimir Peterfeld), A.P., E.B., A.L., DNA Zoo Consortium, S.J.O., A.G.; writing—original draft preparation, A.Y., A.T. (Andrey Tomarovsky), A.T. (Azamat Totikov), S.K., K.-P.K. and T.B.; writing—review and editing, A.Y., A.T. (Andrey Tomarovsky), V.B., P.L.P., S.K., A.G., K.-P.K. and T.B.; visualization, A.Y. and S.K.; supervision, S.K., S.J.O., A.L. and A.G.; project administration, S.K.; funding acquisition, S.K., DNA Zoo Consortium, S.J.O., A.L. and A.G. All authors have read and agreed to the published version of the manuscript.

Funding: The work was supported by a research grant of the Russian Science Foundation (RSF, 19-14-00034-P) and partly by a research grant of the Russian Foundation for Basic Research (RFBR), project number 20-04-00808. A.L. and M.R. were supported by St. Petersburg State University (grant ID PURE: 73023672). N.C. and G.T. were supported by Peter the Great St. Petersburg Polytechnic University in the framework of Russian Federation's Priority 2030 Strategic Academic Leadership Programme (Agreement 75-15-2021-1333). V.P. and E.B. were funded from the state assignment of Ministry of Science and Higher Education of the Russian Federation for Research Centre for Medical Genetics.

Institutional Review Board Statement: Not applicable. We used cultures from the collection of Institute of Molecular and Cellular Biology SB RAS. They were previously described by Beklemisheva et al. 2016, 2020.

Informed Consent Statement: Not applicable.

Data Availability Statement: De novo assembly of male Baikal seal and related reads are available from BioProject PRJNA905417. Hi-C data is available from DNA Zoo SRA BioProject PRJNA512907. Corresponding SRA accessions are listed in Supplementary Table S1. Resequencing data for female seal are available from BioProject PRJNA905443 under SRR22409450 accession. Interactive Hi-C contact maps for the Baikal seal are available at dnazoo.org/assemblies/Pusa_sibirica.

Acknowledgments: We are grateful to Olga Dudchenko, Erez Aiden and Ruqayya Khan from the DNazoo Consortium (Baylor College of Medicine, Houston, TX, USA) for their brilliant help with Hi-C data generation and scaffolding of the Baikal seal genome assembly. We acknowledge Andrei Varnavsky from the Baikal Branch of State Research and the Industrial Center of Fisheries for help with seal sample gathering. We dedicate this paper to the memory of Mikhail Grachev (Limnological Institute, Irkutsk), who discovered Lake Baikal for modern genomic research, including the Baikal seal genome project. Unpublished genome assemblies and sequencing data for cheetah (*Acinonyx jubatus*), mountain lion (*Puma concolor*), domestic dog (*Canis lupus familiaris*), brown bear (*Ursus arctos*), red panda (*Ailurus fulgens*), walrus (*Odobenus rosmarus*), bearded seal (*Erignathus barbatus*), harbor seal (*Phoca vitulina*), spotted seal (*Phoca largha*), and gray seal (*Halichoerus grypus*) are used with permission from the DNA Zoo Consortium (<https://www.dnazoo.org/> (accessed on 21 December 2022)).

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

MP	<u>M</u> aximum <u>p</u> arsimony
ML	<u>M</u> aximum <u>l</u> ikelihood
NJ	<u>N</u> eighbor <u>j</u> oining
BI	<u>B</u> ayesian <u>i</u> nference

References

- Astakhov, V. Middle Pleistocene Glaciations of the Russian North. *Quat. Sci. Rev.* **2004**, *23*, 1285–1311. [[CrossRef](#)]
- Palo, J.U.; Väinölä, R. The Enigma of the Landlocked Baikal and Caspian Seals Addressed through Phylogeny of Phocine Mitochondrial Sequences. *Biol. J. Linn. Soc.* **2006**, *88*, 61–72. [[CrossRef](#)]
- Hassanin, A.; Veron, G.; Ropiquet, A.; Jansen van Vuuren, B.; Lécuyer, A.; Goodman, S.M.; Haider, J.; Nguyen, T.T. Evolutionary History of Carnivora (Mammalia, Laurasiatheria) Inferred from Mitochondrial Genomes. *PLoS ONE* **2021**, *16*, e0240770. [[CrossRef](#)]
- Sasaki, H.; Numachi, K.; Grachev, M.A. The Origin and Genetic Relationships of the Baikal Seal, *Phoca sibirica*, by Restriction Analysis of Mitochondrial DNA. *Zool. Sci.* **2003**, *20*, 1417–1422. [[CrossRef](#)] [[PubMed](#)]
- Petrov, E.A.; Kupchinsky, A.B. Baikal Museum SB RAS Introduction of the Baikal Seal (*Pusa sibirica* Gm.) to Lake Baikal and Current Threats to the Population due to Climate Change: A Review. 1. Introduction of the Baikal Seal in the Lake Baikal: Family Ties, Time and Migration. *Bull. Irkutsk State Univ. Ser. Biol. Ecol.* **2021**, *38*, 103–134. [[CrossRef](#)]
- Miyazaki, N. Caspian and Baikal Seals: *Pusa Caspica* and *Pusa Sibirica*. In *Encyclopedia of Marine Mammals*, 2nd ed.; Academic Press: Cambridge, MA, USA, 2009; pp. 188–191. [[CrossRef](#)]
- Nomokonova, T.; Losey, R.; Iakunaeva, V.; Emel'ianova, I.; Baginova, E.; Pastukhov, M. People and Seals at Siberia's Lake Baikal. *J. Ethnobiol.* **2013**, *33*, 259–280. [[CrossRef](#)]
- Nomokonova, T.; Losey, R.J.; Goriunova, O.I.; Novikov, A.G.; Weber, A.W. A 9000 Year History of Seal Hunting on Lake Baikal, Siberia: The Zooarchaeology of Sagan-Zaba II. *PLoS ONE* **2015**, *10*, e0128314. [[CrossRef](#)]
- Krafft, B.; Kovacs, K.; Lydersen, C. Distribution of Sex and Age Groups of Ringed Seals *Pusa hispida* in the Fast-Ice Breeding Habitat of Kongsfjorden, Svalbard. *Mar. Ecol.-Prog. Ser.* **2007**, *335*, 199–206. [[CrossRef](#)]
- Smith, T.; Stirling, I. The Breeding Habitat of the Ringed Seal (*Phoca hispida*). The Birth Lair and Associated Structures. *Can. J. Zool.* **2011**, *53*, 1297–1305. [[CrossRef](#)]
- Wilson, S.; Dolgova, E.; Trukhanova, I.; Dmitrieva, L.; Crawford, I.; Baimukanov, M.; Goodman, S. Breeding Behavior and Pup Development of the Caspian Seal, *Pusa Caspica*. *J. Mammal.* **2016**, *98*, 143–153. [[CrossRef](#)]
- IWC Commercial Whaling. Available online: <https://iwc.int/management-and-conservation/whaling/commercial> (accessed on 23 January 2023).
- Ozersky, T.; Pastukhov, M.; Poste, A.; Deng, X.; Moore, M. Long-Term and Ontogenetic Patterns of Heavy Metal Contamination in Lake Baikal Seals (*Pusa sibirica*). *Environ. Sci. Technol.* **2017**, *51*, 18. [[CrossRef](#)] [[PubMed](#)]
- Weber, A.; Goriunova, O.I.; Konopatskii, A.K. Prehistoric Seal Hunting on Lake Baikal: Methodology and Preliminary Results of the Analysis of Canine Sections. *J. Archaeol. Sci.* **1993**, *20*, 629–644. [[CrossRef](#)]
- Watanabe, Y.Y.; Baranov, E.A.; Miyazaki, N. Ultrahigh Foraging Rates of Baikal Seals Make Tiny Endemic Amphipods Profitable in Lake Baikal. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 31242–31248. [[CrossRef](#)] [[PubMed](#)]
- Stewart, B.S.; Petrov, E.A.; Baranov, E.A.; Ivanov, A.T.M. Seasonal Movements and Dive Patterns of Juvenile Baikal Seals, *Phoca sibirica*. *Mar. Mammal Sci.* **1996**, *12*, 528–542. [[CrossRef](#)]
- Watanabe, Y.; Baranov, E.A.; Sato, K.; Naito, Y.; Miyazaki, N. Foraging Tactics of Baikal Seals Differ between Day and Night. *Mar. Ecol. Prog. Ser.* **2004**, *279*, 283–289. [[CrossRef](#)]

18. Watanabe, Y.Y.; Baranov, E.A.; Miyazaki, N. Drift Dives and Prolonged Surfacing Periods in Baikal Seals: Resting Strategies in Open Waters? *J. Exp. Biol.* **2015**, *218*, 2793–2798. [[CrossRef](#)]
19. Goodman, S. IUCN Red List of Threatened Species: Pusa Sibirica. 2015. Available online: <https://dx.doi.org/10.2305/IUCN.UK.2016-1.RLTS.T41676A45231738.en> (accessed on 21 December 2022).
20. BaikalNIRO BaikalNIRO—Scientists Assessed the Offspring of the Baikal Seal. Available online: <http://baikal.vniro.ru/ru/novosti/item/104-uchjonye-proveli-otsenku-priploda-bajkal-skoj-nerpy> (accessed on 21 January 2023).
21. Ivanov, K.M.; Kupchinskii, A.B.; Ovdin, M.Y.; Petrov, Y.A.; Sirovatskii, A.A.; Shabanov, D.Y. Experience in application of UAV in ecological studies of the Baikal seal (*Pusa Sibirica* GM.) population at the beginning of shore rookeries formation. *Международный Научно-Исследовательский Журнал* **2022**, *8*, 122. [[CrossRef](#)]
22. Klimova, A.; Phillips, C.D.; Fietz, K.; Olsen, M.T.; Harwood, J.; Amos, W.; Hoffman, J.I. Global Population Structure and Demographic History of the Grey Seal. *Mol. Ecol.* **2014**, *23*, 3999–4017. [[CrossRef](#)]
23. Gilpin, J.B.; Science, N.S.I. of N. On the Seals of Nova Scotia. 1874. Available online: <https://dalspace.library.dal.ca/bitstream/handle/10222/11271/Pt4Art7.pdf?...1> (accessed on 21 December 2022).
24. NOAA Fisheries Gray Seal | NOAA Fisheries. Available online: <https://www.fisheries.noaa.gov/species/gray-seal> (accessed on 22 January 2023).
25. Wood, S.; Frasier, T.; Frasier, B.; Gilbert, J.; White, B.; Bowen, W.; Hammill, M.; Waring, G.; Brault, S. The Genetics of Recolonization: An Analysis of the Stock Structure of Grey Seals (*Halichoerus Grypus*) in the Northwest Atlantic. *Can. J. Zool.* **2011**, *89*, 490–497. [[CrossRef](#)]
26. Han, J.-B.; Sun, F.-Y.; Gao, X.-G.; He, C.-B.; Wang, P.-L.; Ma, Z.-Q.; Wang, Z.-H. Low Microsatellite Variation in Spotted Seal (*Phoca Largha*) Shows a Decrease in Population Size in the Liaodong Gulf Colony. *Ann. Zool. Fenn.* **2010**, *47*, 15–27. [[CrossRef](#)]
27. Conn, P.B.; Ver Hoef, J.M.; McClintock, B.T.; Moreland, E.E.; London, J.M.; Cameron, M.F.; Dahle, S.P.; Boveng, P.L. Estimating Multispecies Abundance Using Automated Detection Systems: Ice-Associated Seals in the Bering Sea. *Methods Ecol. Evol.* **2014**, *5*, 1280–1293. [[CrossRef](#)]
28. Allen, P.J.; Amos, W.; Pomeroy, P.P.; Twiss, S.D. Microsatellite Variation in Grey Seals (*Halichoerus Grypus*) Shows Evidence of Genetic Differentiation between Two British Breeding Colonies. *Mol. Ecol.* **1995**, *4*, 653–662. [[CrossRef](#)] [[PubMed](#)]
29. Boskovic, R.; Kovacs, K.M.; Hammill, M.O.; White, B.N. Geographic Distribution of Mitochondrial DNA Haplotypes in Grey Seals (*Halichoerus Grypus*). *Can. J. Zool.* **1996**, *74*, 1787–1796. [[CrossRef](#)]
30. Nakagawa, E.; Kobayashi, M.; Suzuki, M.; Tsubota, T. Genetic Variation in the Harbor Seal (*Phoca Vitulina*) and Spotted Seal (*Phoca Largha*) Around Hokkaido, Japan, Based on Mitochondrial Cytochrome b Sequences. *Zool. Sci.* **2010**, *27*, 263–268. [[CrossRef](#)] [[PubMed](#)]
31. Fulton, T.L.; Strobeck, C. Multiple Fossil Calibrations, Nuclear Loci and Mitochondrial Genomes Provide New Insight into Biogeography and Divergence Timing for True Seals (Phocidae, Pinnipedia). *J. Biogeogr.* **2010**, *37*, 814–829. [[CrossRef](#)]
32. Fulton, T.L.; Strobeck, C. Multiple Markers and Multiple Individuals Refine True Seal Phylogeny and Bring Molecules and Morphology Back in Line. *Proc. Biol. Sci.* **2010**, *277*, 1065–1070. [[CrossRef](#)]
33. Brandies, P.; Peel, E.; Hogg, C.J.; Belov, K. The Value of Reference Genomes in the Conservation of Threatened Species. *Genes* **2019**, *10*, 846. [[CrossRef](#)]
34. Formenti, G.; Theissinger, K.; Fernandes, C.; Bista, I.; Bombarely, A.; Bleidorn, C.; Ciofi, C.; Crottini, A.; Godoy, J.A.; Höglund, J.; et al. The Era of Reference Genomes in Conservation Genomics. *Trends Ecol. Evol.* **2022**, *37*, 197–202. [[CrossRef](#)]
35. Totikov, A.; Tomarovskiy, A.; Prokopov, D.; Yakupova, A.; Bulyonkova, T.; Derezanin, L.; Rasskazov, D.; Wolfsberger, W.W.; Koepfli, K.-P.; Oleksyk, T.K.; et al. Chromosome-Level Genome Assemblies Expand Capabilities of Genomics for Conservation Biology. *Genes* **2021**, *12*, 1336. [[CrossRef](#)]
36. Paez, S.; Kraus, R.H.S.; Shapiro, B.; Gilbert, M.T.P.; Jarvis, E.D.; Vertebrate Genomes Project Conservation Group; Al-Ajli, F.O.; Ceballos, G.; Crawford, A.J.; Fedrigo, O.; et al. Reference Genomes for Conservation. *Science* **2022**, *377*, 364–366. [[CrossRef](#)]
37. Beklemisheva, V.R.; Perelman, P.L.; Lemskaya, N.A.; Kulemzina, A.I.; Proskuryakova, A.A.; Burkanov, V.N.; Graphodatsky, A.S. The Ancestral Carnivore Karyotype As Substantiated by Comparative Chromosome Painting of Three Pinnipeds, the Walrus, the Steller Sea Lion and the Baikal Seal (Pinnipedia, Carnivora). *PLoS ONE* **2016**, *11*, e0147647. [[CrossRef](#)]
38. Andrews, S. FASTQC. A Quality Control Tool for High Throughput Sequence Data 2010. Available online: <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/> (accessed on 21 December 2022).
39. Marçais, G.; Kingsford, C. A Fast, Lock-Free Approach for Efficient Parallel Counting of Occurrences of k-Mers. *Bioinformatics* **2011**, *27*, 764–770. [[CrossRef](#)]
40. Bolger, A.M.; Lohse, M.; Usadel, B. Trimmomatic: A Flexible Trimmer for Illumina Sequence Data. *Bioinformatics* **2014**, *30*, 2114–2120. [[CrossRef](#)]
41. Leggett, R.M.; Clavijo, B.J.; Clissold, L.; Clark, M.D.; Caccamo, M. NextClip: An Analysis and Read Preparation Tool for Nextera Long Mate Pair Libraries. *Bioinformatics* **2014**, *30*, 566–568. [[CrossRef](#)]
42. Kajitani, R.; Toshimoto, K.; Noguchi, H.; Toyoda, A.; Ogura, Y.; Okuno, M.; Yabana, M.; Harada, M.; Nagayasu, E.; Maruyama, H.; et al. Efficient de Novo Assembly of Highly Heterozygous Genomes from Whole-Genome Shotgun Short Reads. *Genome Res.* **2014**, *24*, 1384–1395. [[CrossRef](#)]

43. Dudchenko, O.; Batra, S.S.; Omer, A.D.; Nyquist, S.K.; Hoeger, M.; Durand, N.C.; Shamim, M.S.; Machol, I.; Lander, E.S.; Aiden, A.P.; et al. De Novo Assembly of the *Aedes Aegypti* Genome Using Hi-C Yields Chromosome-Length Scaffolds. *Science* **2017**, *356*, 92–95. [CrossRef]
44. Dudchenko, O.; Shamim, M.S.; Batra, S.S.; Durand, N.C.; Musial, N.T.; Mostofa, R.; Pham, M.; Glenn St Hilaire, B.; Yao, W.; Stamenova, E.; et al. The Juicebox Assembly Tools Module Facilitates *de Novo* Assembly of Mammalian Genomes with Chromosome-Length Scaffolds for under \$1000. *bioRxiv* **2018**, 254797. [CrossRef]
45. Dudchenko, O. SEAL Team. Available online: <https://www.dnazoo.org/post/seal-team> (accessed on 22 November 2022).
46. Neely, B. This Seal Hits the Spot. Available online: <https://www.dnazoo.org/post/this-seal-hits-the-spot> (accessed on 22 November 2022).
47. Neely, B. Sealing the Deal on Our 250th Genome Assembly. Available online: <https://www.dnazoo.org/post/sealing-the-deal-on-our-250th-genome-assembly> (accessed on 22 November 2022).
48. Park, J.Y.; Kim, K.; Sohn, H.; Kim, H.W.; An, Y.-R.; Kang, J.-H.; Kim, E.-M.; Kwak, W.; Lee, C.; Yoo, D.; et al. Deciphering the Evolutionary Signatures of Pinnipeds Using Novel Genome Sequences: The First Genomes of *Phoca largha*, *Callorhinus ursinus*, and *Eumetopias jubatus*. *Sci. Rep.* **2018**, *8*, 16877. [CrossRef] [PubMed]
49. Manni, M.; Berkeley, M.R.; Seppey, M.; Zdobnov, E.M. BUSCO: Assessing Genomic Data Quality and Beyond. *Curr. Protoc.* **2021**, *1*, e323. [CrossRef] [PubMed]
50. Benson, G. Tandem Repeats Finder: A Program to Analyze DNA Sequences. *Nucleic Acids Res.* **1999**, *27*, 573–580. [CrossRef]
51. Morgulis, A.; Gertz, E.M.; Schäffer, A.A.; Agarwala, R. WindowMasker: Window-Based Masker for Sequenced Genomes. *Bioinformatics* **2006**, *22*, 134–141. [CrossRef] [PubMed]
52. Smit, A.; Hubley, R.; Green, P. RepeatMasker Open-4.0 2013. Available online: <http://www.repeatmasker.org/> (accessed on 21 December 2022).
53. Armstrong, J.; Hickey, G.; Diekhans, M.; Fiddes, I.T.; Novak, A.M.; Deran, A.; Fang, Q.; Xie, D.; Feng, S.; Stiller, J.; et al. Progressive Cactus Is a Multiple-Genome Aligner for the Thousand-Genome Era. *Nature* **2020**, *587*, 246–251. [CrossRef] [PubMed]
54. Krasheninnikova, K.; Diekhans, M.; Armstrong, J.; Dievskii, A.; Paten, B.; O'Brien, S. HalSynteny: A Fast, Easy-to-Use Conserved Synteny Block Construction Method for Multiple Whole-Genome Alignments. *GigaScience* **2020**, *9*, g1aa047. [CrossRef]
55. Arnason, U. Comparative Chromosome Studies in Pinnipedia. *Hereditas* **1974**, *76*, 179–226. [CrossRef]
56. Beklemisheva, V.R.; Perelman, P.L.; Lemskaya, N.A.; Proskuryakova, A.A.; Serdyukova, N.A.; Burkanov, V.N.; Gorshunov, M.B.; Ryder, O.; Thompson, M.; Lento, G.; et al. Karyotype Evolution in 10 Pinniped Species: Variability of Heterochromatin versus High Conservatism of Euchromatin as Revealed by Comparative Molecular Cytogenetics. *Genes* **2020**, *11*, 1485. [CrossRef]
57. Frönicke, L.; Müller-Navia, J.; Romanakis, K.; Scherthan, H. Chromosomal Homeologies between Human, Harbor Seal (*Phoca vitulina*) and the Putative Ancestral Carnivore Karyotype Revealed by Zoo-FISH. *Chromosoma* **1997**, *106*, 108–113. [CrossRef] [PubMed]
58. Graphodatsky, A.; Perelman, P.; O'Brien, S.J. (Eds.) *Atlas of Mammalian Chromosomes*, 2nd ed.; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2020; ISBN 978-1-119-41803-0.
59. Li, H.; Durbin, R. Fast and Accurate Short Read Alignment with Burrows–Wheeler Transform. *Bioinformatics* **2009**, *25*, 1754–1760. [CrossRef]
60. Li, H.; Handsaker, B.; Wysoker, A.; Fennell, T.; Ruan, J.; Homer, N.; Marth, G.; Abecasis, G.; Durbin, R. 1000 Genome Project Data Processing Subgroup The Sequence Alignment/Map Format and SAMtools. *Bioinformatics* **2009**, *25*, 2078–2079. [CrossRef]
61. Pedersen, B.S.; Quinlan, A.R. Mosdepth: Quick Coverage Calculation for Genomes and Exomes. *Bioinformatics* **2018**, *34*, 867–868. [CrossRef]
62. Li, H.; Durbin, R. Inference of Human Population History from Individual Whole-Genome Sequences. *Nature* **2011**, *475*, 493–496. [CrossRef]
63. Cleary, A.C.; Hoffman, J.I.; Forcada, J.; Lydersen, C.; Lowther, A.D.; Kovacs, K.M. 50,000 Years of Ice and Seals: Impacts of the Last Glacial Maximum on Antarctic Fur Seals. *Ecol. Evol.* **2021**, *11*, 14003–14011. [CrossRef]
64. Nikolic, N.; Thompson, P.; de Bruyn, M.; Macé, M.; Chevalet, C. Evolutionary History of a Scottish Harbour Seal Population. *PeerJ* **2020**, *8*, e9167. [CrossRef]
65. Nyman, T.; Valtonen, M.; Aspi, J.; Ruokonen, M.; Kunnasranta, M.; Palo, J.U. Demographic Histories and Genetic Diversities of Fennoscandian Marine and Landlocked Ringed Seal Subspecies. *Ecol. Evol.* **2014**, *4*, 3420–3434. [CrossRef] [PubMed]
66. Paijmans, A.J.; Stoffel, M.A.; Bester, M.N.; Cleary, A.C.; De Bruyn, P.J.N.; Forcada, J.; Goebel, M.E.; Goldsworthy, S.D.; Guinet, C.; Lydersen, C.; et al. The Genetic Legacy of Extreme Exploitation in a Polar Vertebrate. *Sci. Rep.* **2020**, *10*, 5089. [CrossRef] [PubMed]
67. Palo, J.U.; Mäkinen, H.S.; Helle, E.; Stenman, O.; Väinölä, R. Microsatellite Variation in Ringed Seals (*Phoca hispida*): Genetic Structure and History of the Baltic Sea Population. *Heredity* **2001**, *86*, 609–617. [CrossRef] [PubMed]
68. Hoffman, J.I.; Chen, R.S.; Vendrami, D.L.J.; Paijmans, A.J.; Dasmahapatra, K.K.; Forcada, J. Demographic Reconstruction of Antarctic Fur Seals Supports the Krill Surplus Hypothesis. *Genes* **2022**, *13*, 541. [CrossRef]
69. Peart, C.R.; Tusso, S.; Pophaly, S.D.; Botero-Castro, F.; Wu, C.-C.; Auriolles-Gamboa, D.; Baird, A.B.; Bickham, J.W.; Forcada, J.; Galimberti, F.; et al. Determinants of Genetic Variation across Eco-Evolutionary Scales in Pinnipeds. *Nat. Ecol. Evol.* **2020**, *4*, 1095–1104. [CrossRef]

70. Zappes, I.A.; Fabiani, A.; Sbordoni, V.; Rakaj, A.; Palozzi, R.; Allegrucci, G. New Data on Weddell Seal (*Leptonychotes Weddellii*) Colonies: A Genetic Analysis of a Top Predator from the Ross Sea, Antarctica. *PLoS ONE* **2017**, *12*, e0182922. [[CrossRef](#)]
71. Bowen, D. IUCN Red List of Threatened Species: *Halichoerus Grypus*. 2016. Available online: <https://dx.doi.org/10.2305/IUCN.UK.2016-1.RLTS.T9660A45226042.en> (accessed on 21 December 2022).
72. Lowry, L. IUCN Red List of Threatened Species: *Phoca Vitulina*. 2016. Available online: <https://dx.doi.org/10.2305/IUCN.UK.2016-1.RLTS.T17013A45229114.en> (accessed on 21 December 2022).
73. Lowry, L. IUCN Red List of Threatened Species: *Pusa Hispidia*. 2016. Available online: <https://dx.doi.org/10.2305/IUCN.UK.2016-1.RLTS.T41672A45231341.en> (accessed on 21 December 2022).
74. Dobrynin, P.; Liu, S.; Tamazian, G.; Xiong, Z.; Yurchenko, A.A.; Krasheninnikova, K.; Kliver, S.; Schmidt-Küntzel, A.; Koepfli, K.-P.; Johnson, W.; et al. Genomic Legacy of the African Cheetah, *Acinonyx Jubatus*. *Genome Biol.* **2015**, *16*, 277. [[CrossRef](#)]
75. Puma Concolor Isolate SC36_Marlon, Whole Genome Shotgun Sequencing Project 2018. Available online: <https://www.ncbi.nlm.nih.gov/bioproject/PRJNA437459/> (accessed on 21 December 2022).
76. Dudchenko, O. A Study in Scarlet. Available online: <https://www.dnazoo.org/post/a-study-in-scarlet> (accessed on 4 November 2022).
77. Taylor, G.A.; Kirk, H.; Coombe, L.; Jackman, S.D.; Chu, J.; Tse, K.; Cheng, D.; Chuah, E.; Pandoh, P.; Carlsen, R.; et al. The Genome of the North American Brown Bear or Grizzly: *Ursus Arctos Ssp. Horribilis*. *Genes* **2018**, *9*, 598. [[CrossRef](#)]
78. Hu, Y.; Wu, Q.; Ma, S.; Ma, T.; Shan, L.; Wang, X.; Nie, Y.; Ning, Z.; Yan, L.; Xiu, Y.; et al. Comparative Genomics Reveals Convergent Evolution between the Bamboo-Eating Giant and Red Pandas. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 1081–1086. [[CrossRef](#)] [[PubMed](#)]
79. Foote, A.D.; Liu, Y.; Thomas, G.W.C.; Vinař, T.; Alföldi, J.; Deng, J.; Dugan, S.; van Elk, C.E.; Hunter, M.E.; Joshi, V.; et al. Convergent Evolution of the Genomes of Marine Mammals. *Nat. Genet.* **2015**, *47*, 272–275. [[CrossRef](#)] [[PubMed](#)]
80. Clavijo, B.J.; Venturini, L.; Schudoma, C.; Accinelli, G.G.; Kaithakottil, G.; Wright, J.; Borrill, P.; Kettleborough, G.; Heavens, D.; Chapman, H.; et al. An Improved Assembly and Annotation of the Allohexaploid Wheat Genome Identifies Complete Families of Agronomic Genes and Provides Genomic Evidence for Chromosomal Translocations. *Genome Res.* **2017**, *27*, 885–896. [[CrossRef](#)]
81. *Phoca Vitulina* Isolate PV1807, Whole Genome Shotgun Sequencing Project 2019. Available online: <https://www.ncbi.nlm.nih.gov/bioproject/494181> (accessed on 21 December 2022).
82. Löytynoja, A.; Goldman, N. Phylogeny-Aware Gap Placement Prevents Errors in Sequence Alignment and Evolutionary Analysis. *Science* **2008**, *320*, 1632–1635. [[CrossRef](#)]
83. Castresana, J. Selection of Conserved Blocks from Multiple Alignments for Their Use in Phylogenetic Analysis. *Mol. Biol. Evol.* **2000**, *17*, 540–552. [[CrossRef](#)] [[PubMed](#)]
84. Nguyen, L.-T.; Schmidt, H.A.; von Haeseler, A.; Minh, B.Q. IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Mol. Biol. Evol.* **2015**, *32*, 268–274. [[CrossRef](#)]
85. Kalyaanamoorthy, S.; Minh, B.Q.; Wong, T.K.F.; von Haeseler, A.; Jermini, L.S. ModelFinder: Fast Model Selection for Accurate Phylogenetic Estimates. *Nat. Methods* **2017**, *14*, 587–589. [[CrossRef](#)]
86. Yang, Z. PAML 4: Phylogenetic Analysis by Maximum Likelihood. *Mol. Biol. Evol.* **2007**, *24*, 1586–1591. [[CrossRef](#)]
87. Brashear, W.A.; Bredemeyer, K.R.; Murphy, W.J. Genomic Architecture Constrained Placental Mammal X Chromosome Evolution. *Genome Res.* **2021**, *31*, 1353–1365. [[CrossRef](#)]
88. Lewin, H.A.; Graves, J.A.M.; Ryder, O.A.; Graphodatsky, A.S.; O'Brien, S.J. Precision Nomenclature for the New Genomics. *GigaScience* **2019**, *8*, giz086. [[CrossRef](#)]
89. Dyomin, A.G.; Koshel, E.I.; Kiselev, A.M.; Saifitdinova, A.F.; Galkina, S.A.; Fukagawa, T.; Kostareva, A.A.; Gaginskaya, E.R. Chicken RRNA Gene Cluster Structure. *PLoS ONE* **2016**, *11*, e0157464. [[CrossRef](#)] [[PubMed](#)]
90. Levan, A.; Fredga, K.; Sandberg, A.A. Nomenclature for Centromeric Position on Chromosomes. *Hereditas* **1964**, *52*, 201–220. [[CrossRef](#)]
91. McGowan-Jordan, J.; Hastings, R.J.; Moore, S. *ISCN 2020: An International System for Human Cytogenomic Nomenclature (2020)*, 1st ed.; S. Karger: Basel, Switzerland, 2020; ISBN 978-3-318-06706-4.
92. Huynh, L.Y.; Maney, D.L.; Thomas, J.W. Chromosome-Wide Linkage Disequilibrium Caused by an Inversion Polymorphism in the White-Throated Sparrow (*Zonotrichia Albicollis*). *Heredity* **2011**, *106*, 537–546. [[CrossRef](#)] [[PubMed](#)]
93. Kennington, W.J.; Partridge, L.; Hoffmann, A.A. Patterns of Diversity and Linkage Disequilibrium Within the Cosmopolitan Inversion In(3R)Payne in *Drosophila Melanogaster* Are Indicative of Coadaptation. *Genetics* **2006**, *172*, 1655–1663. [[CrossRef](#)]
94. Ayala, D.; Guerrero, R.F.; Kirkpatrick, M. Reproductive isolation and local adaptation quantified for a chromosome inversion in a malaria mosquito. *Evolution* **2013**, *67*, 946–958. [[CrossRef](#)]
95. Cursino, M.S.; Salviano, M.B.; Abril, V.V.; dos Santos Zanetti, E.; Duarte, J.M.B. The Role of Chromosome Variation in the Speciation of the Red Brocket Deer Complex: The Study of Reproductive Isolation in Females. *BMC Evol. Biol.* **2014**, *14*, 40. [[CrossRef](#)]
96. Kempainen, P.; Knight, C.G.; Sarma, D.K.; Hlaing, T.; Prakash, A.; Maung Maung, Y.N.; Somboon, P.; Mahanta, J.; Walton, C. Linkage Disequilibrium Network Analysis (LDna) Gives a Global View of Chromosomal Inversions, Local Adaptation and Geographic Structure. *Mol. Ecol. Resour.* **2015**, *15*, 1031–1045. [[CrossRef](#)]

97. Noor, M.A.F.; Grams, K.L.; Bertucci, L.A.; Reiland, J. Chromosomal Inversions and the Reproductive Isolation of Species. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 12084–12088. [[CrossRef](#)]
98. Marquès-Bonet, T.; Cáceres, M.; Bertranpetit, J.; Preuss, T.M.; Thomas, J.W.; Navarro, A. Chromosomal Rearrangements and the Genomic Distribution of Gene-Expression Divergence in Humans and Chimpanzees. *Trends Genet. TIG* **2004**, *20*, 524–529. [[CrossRef](#)]
99. Arnason, U.; Gullberg, A.; Janke, A.; Kullberg, M.; Lehman, N.; Petrov, E.A.; Väinölä, R. Pinniped Phylogeny and a New Hypothesis for Their Origin and Dispersal. *Mol. Phylogenet. Evol.* **2006**, *41*, 345–354. [[CrossRef](#)] [[PubMed](#)]
100. Stoffel, M.A.; Humble, E.; Paijmans, A.J.; Acevedo-Whitehouse, K.; Chilvers, B.L.; Dickerson, B.; Galimberti, F.; Gemmell, N.J.; Goldsworthy, S.D.; Nichols, H.J.; et al. Demographic Histories and Genetic Diversity across Pinnipeds Are Shaped by Human Exploitation, Ecology and Life-History. *Nat. Commun.* **2018**, *9*, 4836. [[CrossRef](#)] [[PubMed](#)]
101. Willi, Y.; Kristensen, T.N.; Sgrò, C.M.; Weeks, A.R.; Ørsted, M.; Hoffmann, A.A. Conservation Genetics as a Management Tool: The Five Best-Supported Paradigms to Assist the Management of Threatened Species. *Proc. Natl. Acad. Sci. USA* **2022**, *119*, e2105076119. [[CrossRef](#)] [[PubMed](#)]
102. Venn, O.; Turner, I.; Mathieson, I.; de Groot, N.; Bontrop, R.; McVean, G. Nonhuman Genetics. Strong Male Bias Drives Germline Mutation in Chimpanzees. *Science* **2014**, *344*, 1272–1275. [[CrossRef](#)] [[PubMed](#)]
103. Pacifici, M.; Santini, L.; Marco, M.D.; Baisero, D.; Francucci, L.; Marasini, G.G.; Visconti, P.; Rondinini, C. Generation Length for Mammals. *Nat. Conserv.* **2013**, *5*, 89–94. [[CrossRef](#)]
104. McLaren, I. On the Origin of the Caspian and Baikal Seals and the Paleoclimatological Implication. *Am. J. Sci.* **1960**, *258*, 47–65. [[CrossRef](#)]
105. Fisheries, N. Ringed Seal | NOAA Fisheries. Available online: <https://www.fisheries.noaa.gov/species/ringed-seal> (accessed on 30 January 2023).
106. Clark, P.U.; Archer, D.; Pollard, D.; Blum, J.D.; Rial, J.A.; Brovkin, V.; Mix, A.C.; Pisias, N.G.; Roy, M. The Middle Pleistocene Transition: Characteristics, Mechanisms, and Implications for Long-Term Changes in Atmospheric PCO₂. *Crit. Quat. Stratigr.* **2006**, *25*, 3150–3184. [[CrossRef](#)]
107. Pisias, N.G.; Moore, T.C. The Evolution of Pleistocene Climate: A Time Series Approach. *Earth Planet. Sci. Lett.* **1981**, *52*, 450–458. [[CrossRef](#)]
108. Bajo, P.; Drysdale, R.N.; Woodhead, J.D.; Hellstrom, J.C.; Hodell, D.; Ferretti, P.; Voelker, A.H.L.; Zanchetta, G.; Rodrigues, T.; Wolff, E.; et al. Persistent Influence of Obliquity on Ice Age Terminations since the Middle Pleistocene Transition. *Science* **2020**, *367*, 1235–1239. [[CrossRef](#)]
109. Elderfield, H.; Ferretti, P.; Greaves, M.; Crowhurst, S.; McCave, I.N.; Hodell, D.; Piotrowski, A.M. Evolution of Ocean Temperature and Ice Volume Through the Mid-Pleistocene Climate Transition. *Science* **2012**, *337*, 704–709. [[CrossRef](#)]
110. Clotten, C.; Stein, R.; Fahl, K.; Schreck, M.; Risebrobakken, B.; De Schepper, S. On the Causes of Arctic Sea Ice in the Warm Early Pliocene. *Sci. Rep.* **2019**, *9*, 989. [[CrossRef](#)]
111. Dolan, A.M.; Haywood, A.M.; Hunter, S.J.; Tindall, J.C.; Dowsett, H.J.; Hill, D.J.; Pickering, S.J. Modelling the Enigmatic Late Pliocene Glacial Event—Marine Isotope Stage M2. *Glob. Planet. Chang.* **2015**, *128*, 47–60. [[CrossRef](#)]
112. Knies, J.; Matthiessen, J.; Vogt, C.; Laberg, J.S.; Hjelstuen, B.O.; Smelror, M.; Larsen, E.; Andreassen, K.; Eidvin, T.; Vorren, T.O. The Plio-Pleistocene Glaciation of the Barents Sea–Svalbard Region: A New Model Based on Revised Chronostratigraphy. *Quat. Sci. Rev.* **2009**, *28*, 812–829. [[CrossRef](#)]
113. Butt, F.A.; Elverhøi, A.; Solheim, A.; Forsberg, C. Deciphering Late Cenozoic Development of the Western Svalbard Margin from ODP Site 986 Results. *Mar. Geol.* **2000**, *169*, 373–390. [[CrossRef](#)]
114. Sejrup, H.P.; Hjelstuen, B.O.; Torbjørn Dahlgren, K.L.; Hafliðason, H.; Kuijpers, A.; Nygård, A.; Praeg, D.; Stoker, M.S.; Vorren, T.O. Pleistocene Glacial History of the NW European Continental Margin. *Mar. Pet. Geol.* **2005**, *22*, 1111–1129. [[CrossRef](#)]
115. Solheim, A.; Faleide, J.I.; Andersen, E.S.; Elverhøi, A.; Forsberg, C.F.; Vanneste, K.; Uenzelmann-neben, G.; Channell, J.E.T. Late cenozoic seismic stratigraphy and glacial geological development of the east greenland and svalbard–barents sea continental margins. *Quat. Sci. Rev.* **1998**, *17*, 155–184. [[CrossRef](#)]
116. Solheim, A.; Andersen, E.S.; Elverhøi, A.; Fiedler, A. Late Cenozoic Depositional History of the Western Svalbard Continental Shelf, Controlled by Subsidence and Climate. *Glob. Planet. Chang.* **1996**, *12*, 135–148. [[CrossRef](#)]
117. Björck, S. A Review of the History of the Baltic Sea, 13.0–8.0 Ka BP. *Quat. Int.* **1995**, *27*, 19–40. [[CrossRef](#)]
118. Tokarev, I.V.; Borodulina, G.S.; Subetto, D.A.; Voronyuk, G.Y.; Zobkov, M.B. Fingerprint of the Geographic and Climate Evolution of the Baltic–White Sea Region in the Late Pleistocene–Holocene in Groundwater Stable Isotopes (2H, 18O). *Quat. Int.* **2019**, *524*, 76–85. [[CrossRef](#)]
119. Näslund, J.-O.; Moren, L.; Bogren, J.; Gustavsson, T.; Moberg, A.; Alexandersson, H.; Jansson, P.; Whitehouse, P.; Hartikainen, J.; Wallroth, T.; et al. *Climate and Climate Related Issues for the Safety Assessment SR-Can*; Svensk Kärnbränslehantering AB: Stockholm, Sweden, 2006.
120. Krinner, G.; Mangerud, J.; Jakobsson, M.; Crucifix, M.; Ritz, C.; Svendsen, J.I. Enhanced Ice Sheet Growth in Eurasia Owing to Adjacent Ice-Dammed Lakes. *Nature* **2004**, *427*, 429–432. [[CrossRef](#)]

121. Dahl-Jensen, D.; Albert, M.R.; Aldahan, A.; Azuma, N.; Balslev-Clausen, D.; Baumgartner, M.; Berggren, A.-M.; Bigler, M.; Binder, T.; Blunier, T.; et al. Eemian Interglacial Reconstructed from a Greenland Folded Ice Core. *Nature* **2013**, *493*, 489–494. [[CrossRef](#)]
122. Lambeck, K.; Nakada, M. Constraints on the Age and Duration of the Last Interglacial Period and on Sea-Level Variations. *Nature* **1992**, *357*, 125–128. [[CrossRef](#)]
123. Severinghaus, J.P.; Brook, E.J. Abrupt Climate Change at the End of the Last Glacial Period Inferred from Trapped Air in Polar Ice. *Science* **1999**, *286*, 930–934. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.