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Progress of Mesenchymal Stem Cell Therapy for Neural and Retinal Diseases

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WJSC 6th Anniversary Special Issues (2): Mesenchymal stem cells**Progress of mesenchymal stem cell therapy for neural and retinal diseases**

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Abstract

Complex circuitry and limited regenerative power make central nervous system (CNS) disorders the most challenging and difficult for functional repair. With elusive disease mechanisms, traditional surgical and medical interventions merely slow down the progression of the neurodegenerative diseases. However, the number of neurons still diminishes in many patients. Recently, stem cell therapy has been proposed as a viable option. Mesenchymal stem cells (MSCs), a widely-studied human adult stem cell population, have been discovered for more than 20 years. MSCs have been found all over the body and can be conveniently obtained from different accessible tissues: bone marrow, blood, and adipose and dental tissue. MSCs have high proliferative and differentiation abilities, providing an inexhaustible source of neurons and glia for cell replacement therapy. Moreover, MSCs also show neuroprotective effects

without any genetic modification or reprogramming. In addition, the extraordinary immunomodulatory properties of MSCs enable autologous and heterologous transplantation. These qualities heighten the clinical applicability of MSCs when dealing with the pathologies of CNS disorders. Here, we summarize the latest progress of MSC experimental research as well as human clinical trials for neural and retinal diseases. This review article will focus on multiple sclerosis, spinal cord injury, autism, glaucoma, retinitis pigmentosa and age-related macular degeneration.

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Key words: Mesenchymal stem cells; Stem cell therapy; Central nervous system; Retina; Clinical trial

Core tip: Central nervous system (CNS) disorders are the most challenging and difficult for functional repair. Neurons are still diminishing in many patients despite surgical and medical interventions. Stem cell therapy has been proposed as a viable option. Mesenchymal stem cell (MSC) is a widely-studied human adult stem cell population. MSCs can be conveniently obtained from different accessible tissues. MSCs have high proliferative and differentiation abilities, providing an inexhaustible source of neurons and glia. MSCs also show neuroprotective effects and possess extraordinary immunomodulatory properties. These qualities heighten the clinical applicability of MSCs when dealing with the pathologies of CNS disorders.

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STEM CELL THERAPY AND MESENCHYMAL STEM CELLS

Stem cells are undifferentiated cells defined by their ability to self-renew and differentiate into mature cells. Stem cells are attractive because they are highly proliferative, implying that an inexhaustible number of mature cells can be generated from a given stem cell source. On this basis, cell replacement therapy has been proposed in recent years as a viable alternative for various pathologies. Cell replacement therapy hypothesizes that new retinal cells could be generated from stem cells so as to substitute the damaged cells in the diseased retina. This theory is mainly established from embryonic stem cells (ESCs) and induced pluripotent stem cells (iPSCs). In addition to cell replacement function, stem cells could have another protective effect, the paracrine effect. The paracrine effects of stem cells are believed to modulate the micro-environment of the diseased tissues so as to protect the injured cells, promote survival and activate any available endogenous repair mechanisms. This latter observation applies mainly to the transplantation of adult stem cells.

Adult stem cells are defined as the stem cells found in fully developed tissues. The function of adult stem cells is the maintenance of adult tissue specificity by homeostatic cell replacement and tissue regeneration^[1]. Adult stem cells are presumed quiescent within adult tissues, but divide infrequently to maintain their own niche by generating a stem cell clone and a transiently-amplifying cell. The transiently-amplifying cells will undergo a limited number of cell divisions before terminal differentiation into mature functional tissue cells. The existence of adult stem cells has been reported in multiple organs; these include: brain, heart, skin, intestine, testis, muscle and blood, among others.

Mesenchymal stem cells (MSCs), also called marrow stromal cells, are an adult stem cell population of stromal progenitor cells of mesodermal origin^[2]. MSCs were originally identified in the bone marrow, representing 0.001%-0.01% of the bone marrow population. MSCs can also be found in other systems all over the body, such as adipose tissue, liver, umbilical cord, central nervous system (CNS) and dental tissues^[3]. According to the International Society of Cellular Therapy^[4], the minimal criteria to define MSCs are: (1) grown in adherence to plastic surface of dishes when maintained in standard culture conditions; (2) positive expression of cytospecific cell surface markers (CD105, CD90 and CD73) and negative expression of other cell surface markers (CD45, CD34, CD14 and CD11b); and (3) capacity to differentiate into mesenchymal lineages, under appropriate *in vitro* conditions. In addition to the expression of the three cell surface markers, MSCs also express CD29, CD44, CD146 and STRO-1^[5].

The function of MSCs is to differentiate into osteocytes, chondrocytes, myoblasts and adipocytes^[6,7]. An increasing number of studies, however, report that MSCs are capable of giving rise to cells of an entirely distinct

lineage, including neuron-like cells. MSCs are not only able to differentiate into neurons for cell replacement therapy, they also exert paracrine effects by modulating the plasticity of damaged host tissues, secreting neurotrophic and survival-promoting growth factors, restoring synaptic transmitter release, integrating into existing neural and synaptic networks, and re-establishing functional afferent and efferent connections^[8]. These paracrine activities have not been reported in ESCs or iPSCs. Moreover, MSCs possess strong immunosuppressive properties and inhibit the release of pro-inflammatory cytokines^[9]. This allows autologous, as well as, allogeneic transplantation of MSCs without the need of pharmacological immunosuppression. Furthermore, MSCs can be transplanted directly without genetic modification or pre-treatments, and are able to migrate to the tissue injury sites^[10]. In addition, there is no teratoma formation concern after transplantation^[11], and no moral objection or ethical controversies involved in their attainment^[12]. These advantageous properties, as well as the expansion potential of MSCs initiate the idea of clinical applications of MSCs to treat different human diseases, especially CNS disorders. Currently, over 100 MSC clinical trials for different diseases have been listed by the United States National Institutes of Health trial database (www.clinicaltrials.gov), indicating that MSC therapy is a popular trend for the field of regenerative medicine in the years to come.

This review article provides an update on the progress of MSC experimental research as well as human clinical trials for neural and retinal diseases with emphasis on multiple sclerosis, spinal cord injury, autism, glaucoma, retinitis pigmentosa and age-related macular degeneration.

MULTIPLE SCLEROSIS

Multiple sclerosis (MS) is an immune-mediated neurodegenerative disorder of the CNS, affecting over 1.3 million people worldwide. The histopathological hallmark of MS is the formation of an inflammatory plaque, which originates from a breach in the integrity of the blood-brain barrier^[13]. The histologic features of lesions in MS include: lymphocyte infiltration, loss of oligodendrocytes, demyelination, and widespread axonal damage^[14]. Myelin-reactive T cells, which secrete interferon- and interleukins, have been suggested to be responsible for the inflammatory demyelination seen in MS^[15]. Currently, there are three treatment options approved by the Food and Drug Administration (FDA) for MS: administration of interferon beta, glatiramer acetate, or mitoxantrone^[16]. However, there is still no medical cure for MS.

Experimental autoimmune encephalomyelitis (EAE), the best known and most commonly used model for MS, mechanistically defines the immune processes responsible for the clinical manifestations and development of MS^[17]. This animal model provides insight for the application of immunotherapy to treat MS^[18]. MSCs have been pro-

posed as a treatment for autoimmune diseases, including MS, because of their immunosuppressive properties and neural repair function^[19]. Transplantation of human MSCs into animals with ongoing EAE results in rapid and sustained functional recovery due to a reduced number of inflammatory myelin-specific Th1 cells and astrocytes as well as an increased number of inflammatory-inhibiting Th2 cells, oligodendrocytes and neurons^[20]. This functional benefit is a critical stepping-stone towards effective MSC therapies in MS patients.

Among all of the CNS disorders, MS has the highest number of registered clinical trials. Altogether there are 14 registered clinical trials for MS (Table 1), and two of them have been published. The study from Israel is a phase-1/2 open safety clinical trial to evaluate the feasibility, safety and immunological effects of intrathecal and intravenous administration of autologous MSCs in 15 MS patients (NCT00781872; <http://clinicaltrials.gov/>)^[21]. No major adverse effects have been reported in this study, and the mean Expanded Disability Status Scale (EDSS) improved from 6.7 to 5.9 (EDSS steps 1.0 to 4.5: MS patients are fully ambulatory, whereas EDSS steps 5.0 to 9.5: MS patients are impaired to ambulation). Moreover, magnetic resonance imaging visualized the MSCs in the occipital horns of the ventricles, indicating migration of the cells. In addition, the proportion of CD4+/CD25+ regulatory T cells increased, whereas the proliferative responses of lymphocytes decreased. The mesenchymal stem cells in the multiple sclerosis trial (MSCIMS) originated in the United Kingdom, is an open-label phase 2a proof-of-concept study of autologous MSCs in secondary progressive MS (NCT00395200; <http://clinicaltrials.gov/>)^[22,23]. In this study, 10 patients received intravenous infusion of autologous bone marrow-derived MSCs (1.6×10^6 cells per kg body weight). The “sentinel lesion approach” assessing the anterior visual pathway was used to measure the efficacy of treatment. Results show that treatment improved visual acuity, visual evoked response latency, and increased the optic nerve area of the recipients. No serious adverse events were identified. For other clinical trials, mainly autologous MSCs have been used, although one study from China uses umbilical cord MSCs (NCT01364246; <http://clinicaltrials.gov/>). Interestingly, an open-label phase I clinical trial from New York was designed to evaluate autologous MSC-derived neural progenitor cells in progressive MS patients (NCT01933802; <http://clinicaltrials.gov/>) even though neural stem cells from EAE animals mainly develop astrocytes rather than oligodendrocytes, or oligodendrocyte precursor cells and neurons^[20].

SPINAL CORD INJURY

Spinal cord injury (SCI) is the most devastating and traumatic disorder among CNS conditions^[24]. The worldwide frequency of SCI is about 40 cases per million individuals^[25]. SCI can be caused by traffic accidents, violent assaults, falls, sport and other traumatic events. Depending on the injury location, extent, phases and time frames,

SCI therapeutic strategies can vary greatly^[26]. Most SCI patients are in the chronic phase, characterized by ongoing demyelination, local inflammation and apoptosis, decreased number of activated macrophages, and formation of glial scar and pseudocysts^[27]. The present standard treatment for SCI patients is surgical intervention, high doses of methylprednisolone, and symptomatic therapy followed by rehabilitation^[28]. New neuroregenerative strategies will be focused on neuroprotection and axonal regeneration in a permissive environment.

Cellular therapy aims to reconstruct the spinal cord through cellular replacement, glial scar remodeling, axonal guidance, and filling of formed syringomyelia^[29]. *In vivo* administration of MSCs in different SCI animal models showed functional recovery including: increased motor activity and sensation in the paralyzed limbs, reduced cavity formation in the spinal cord, and axonal sprouting through the glial scar^[30,31]. The objective of MSC application is to ameliorate the consequences of secondary injury by preserving the host nerve cells, rather than replacing them^[32].

Comparable to MS studies, there are 11 registered clinical trials using MSCs for SCI treatment (Table 1), among which two studies (one from Egypt and one from South Korea) have been completed. The Korean study investigated the safety of single intravenous infusion of autologous adipose tissue-derived MSCs (4×10^8 cells) in 8 male patients with chronic SCI (NCT01274975; <http://clinicaltrials.gov/>)^[33]. No adverse events were observed. Although one patient showed improvement in the American Spinal Injury Association (ASIA) scale from grade A (No sensory or motor function is preserved in sacral segments S4-S5) to grade C (Motor function is preserved below the neurologic level, and most key muscles below the neurologic level have muscle grade less than 3) and three patients showed motor score improvement, this phase I clinical trial might not have the statistical power to conclude on the efficacy of treatment effect with adipose tissue-derived MSCs on SCI. The study conducted in Egypt (NCT00816803; <http://clinicaltrials.gov/>), is a Phase-1/2 clinical trial applying bone marrow-derived MSCs at the injury site of chronic SCI patients. However, no results of this study have been released. Finally, there are two Phase-3 clinical trials taking place in China (NCT01873547; <http://clinicaltrials.gov/>) and Korea (NCT01676441; <http://clinicaltrials.gov/>). The study in China plans to use umbilical cord MSCs to treat 100 chronic SCI patients compared to the rehabilitation-only group and no stem cell and rehabilitation group, whereas the study in Korea was designed to transplant bone marrow-derived MSCs to treat 32 chronic SCI patients. For other ongoing clinical trials in SCI, the approaches are mainly intrathecal transplantation of bone marrow-derived MSCs and adipose tissue-derived MSCs in chronic SCI patients.

AUTISM

Autism belongs to a spectrum of heterogeneous neuro-

Table 1 Registered clinical trials on mesenchymal stem cells for neural diseases

Identifier	Country	Status	Study	Phase of trial	Estimated number of patients	Estimated trial end	Disease
NCT01377870	Iran	Recruiting	Evaluation of autologous mesenchymal stem cell transplantation (effects and side effects) in multiple sclerosis	Phase 1/2	30	2013	Multiple sclerosis
NCT01895439	Jordan	Recruiting	Safety and efficacy study of autologous bone marrow mesenchymal stem cells in multiple sclerosis	Phase 1/2	30	2014	Multiple sclerosis
NCT01883661	India	Not yet recruiting	Safety and efficacy of MSCs in MS	Phase 1/2	15	2015	Multiple sclerosis
NCT00395200	United Kingdom	Completed	MSCIMS	Phase 1/2	10	2010	Multiple sclerosis
NCT01854957	Italy	Recruiting	MESEMS	Phase 1/2	20	2014	Multiple sclerosis
NCT01730547	Sweden	Recruiting	Mesenchymal stem cells for multiple sclerosis	Phase 1/2	15	2015	Multiple sclerosis
NCT01364246	China	Recruiting	Safety and efficacy of umbilical cord mesenchymal stem cell therapy for patients with progressive multiple sclerosis and neuromyelitis optica	Phase 1/2	20	2014	Multiple sclerosis
NCT01056471	Spain	Unknown	Autologous mesenchymal stem cells from adipose tissue in patients with secondary progressive multiple sclerosis (CMM/EM/2008)	Phase 1/2	30	2012	Multiple sclerosis
NCT01228266	Spain	Active, not recruiting	Mesenchymal stem cell transplantation in MS (CMM-EM)	Phase 2	16	2013	Multiple sclerosis
NCT00813969	United States	Active, not recruiting	Autologous MSC transplantation in MS	Phase 1	24	2014	Multiple sclerosis
NCT01933802	United States	Not yet recruiting	Intrathecal administration of autologous MSC-NP in patients with multiple sclerosis	Phase 1	20	2016	Multiple sclerosis
NCT01606215	United Kingdom	Recruiting	STREAMS	Phase 1/2	13	2015	Multiple sclerosis
NCT01745783	Spain	Recruiting	Mesenchymal cells from autologous bone marrow, administered intravenously in patients diagnosed with multiple sclerosis	Phase 1/2	30	2014	Multiple sclerosis
NCT00781872	Israel	Unknown	MSCs for the treatment of MS	Phase 1/2	20	2009	Multiple sclerosis
NCT01694927	Chile	Enrolling by invitation	Autologous mesenchymal stem cells in spinal cord injury (SCI) patients (MSC-SCI)	Phase 2	30	2014	Spinal cord injury
NCT01446640	China	Recruiting	Mesenchymal stem cells transplantation to patients with spinal cord injury (MSC)	Phase 1/2	20	2014	Spinal cord injury
NCT01676441	South Korea	Recruiting	Safety and efficacy of autologous mesenchymal stem cells in chronic spinal cord injury	Phase 2/3	32	2014	Spinal cord injury
NCT01769872	South Korea	Recruiting	Safety and effect of adipose tissue derived mesenchymal stem cell implantation in patients with spinal cord injury	Phase 1/2	15	2014	Spinal cord injury
NCT01162915	United States	Active, not recruiting	Transfer of bone marrow derived stem cells for the treatment of spinal cord injury	Phase 1	10	2013	Spinal cord injury
NCT01274975	South Korea	Completed	Autologous adipose derived mscs transplantation in patient with spinal cord injury	Phase 1	8	2010	Spinal cord injury
NCT01624779	South Korea	Recruiting	Intrathecal transplantation of autologous adipose tissue derived msc in the patients with spinal cord injury	Phase 1	15	2013	Spinal cord injury
NCT01393977	China	Unknown	Difference between rehabilitation therapy and stem cells transplantation in patients with spinal cord injury in China	Phase 2	60	2012	Spinal cord injury
NCT01873547	China	Recruiting	Different efficacy between rehabilitation therapy and stem cells transplantation in patients with SCI in China (SCI-III)	Phase 3	300	2014	Spinal cord injury
NCT01325103	Brazil	Unknown	Autologous bone marrow stem cell transplantation in patients with spinal cord injury	Phase 1	20	2013	Spinal cord injury
NCT00816803	Egypt	Completed	Cell transplant in spinal cord injury patients	Phase 1/2	80	2008	Spinal cord injury
NCT01343511	China	Completed	Safety and efficacy of stem cell therapy in patients with autism	Phase 1/2	37	2011	Autism

Information obtained from <http://clinicaltrials.gov/>. MSCs: Mesenchymal stem cells; MS: Multiple sclerosis; MSCIMS: Mesenchymal Stem Cells in Multiple Sclerosis; MESEMS: MEsenchymal StEm Cells for Multiple Sclerosis; MSC-NP: Mesenchymal Stem Cell-derived Neural Progenitors; STREAMS: Stem Cells in Rapidly Evolving Active Multiple Sclerosis.

developmental disorders^[34]. It is characterized by abnormalities in social interaction, impaired verbal and nonverbal communication, and repetitive, obsessive behavior^[35]. According to the Centers for Disease Control, the prevalence of autism hovers around 60 in every 10000 children^[36]. Even though there is no defined gold standard approach, current interventions for autism can be divided into behavioral, nutritional and pharmacological^[37]. Medical interventions aim to ameliorate the neuropsychiatric disorders associated with autism. The medications include selective serotonin reuptake inhibitors (SSRI's), typical and atypical anti-psychotic drugs, psycho-stimulants, α -2 agonists, β blockers, lithium, anti-convulsant mood stabilizers and anti-depressants^[38-40]. Unfortunately, autism is still not treatable.

The pathogenic mechanism of autism is not clearly understood and remains elusive. Nevertheless, two pathologies are commonly found within the autism patients: the first observation is an impaired central nervous system circulation and hypoperfusion to the brain, whereas the second observation is systemic T cell and B cell abnormalities as well as active neuroinflammatory processes in the brain^[41]. Based on the immunomodulatory properties of MSCs, therapies employing MSCs have been proposed to target the immune deregulation observed in autism. Basically, it is believed that MSCs are able to inhibit the release of pro-inflammatory cytokines and have strong immunosuppressive activity^[42]. This not only allows for autologous transplantation, but also heterologous transplantation without the requirement of pharmacological immunosuppression^[43].

Currently, there is only one registered human clinical trial using MSCs to treat autism (NCT01343511; <http://www.clinicaltrials.gov/>; Table 1). This study aimed to test the safety and efficacy of human umbilical cord MSCs and human cord blood mononuclear cell transplantation in Chinese patients with autism^[44]. Outcomes from this study assuaged the safety concerns in using MSCs and mononuclear cells for transplantation in autism patients, and no severe adverse effects were observed. In addition, results also showed that combined transplantation of MSCs and mononuclear cells (combination group) had better therapeutic effects than transplantation of mononuclear cells alone (CBMNC group) in terms of the Childhood Autism Rating Scale (CARS) total score (combination group: 28.00 ± 6.18 ; CBMNC group: 37.14 ± 10.15 ; CARS total score > 30 means the child is considered to be autistic), Clinical Global Impression (CGI) scale (combination group: 88% much improved or higher; CBMNC group: 49% much improved or higher) and the Aberrant Behavior Checklist (ABC) total score (combination group: 36.78 ± 16.95 ; CBMNC group: 58.36 ± 31.73 ; a high score indicates greater severity while a low score indicates a milder degree of difficulty).

GLAUCOMA

Glaucoma is a group of chronic, degenerative optic

neuropathies. It is characterized by a slow progressive degeneration of retinal ganglion cells (RGCs) and their axons, which results in visual field defects^[45]. Glaucoma is the leading cause of irreversible blindness, affecting more than 60 million people worldwide^[46]. Traditional and current treatments for glaucoma are based on surgical or medical interventions to slow disease progression and limit visual loss^[47]. However, in many patients, the numbers of RGCs still diminish, and glaucoma cannot be completely cured.

The molecular basis of glaucoma is complex. The pathophysiological mechanisms leading to RGC degeneration in glaucoma include a complex interaction between primary axonal injury, neurotrophic factor deprivation, ischemia, oxidative stress, mitochondrial dysfunction and inflammation^[48]. New therapies aim to supplement neurotrophic factors, such as brain-derived neurotrophic factor (BDNF), ciliary neurotrophic factor (CNTF), glial cell line-derived neurotrophic factor (GDNF) and nerve growth factor (NGF)^[49]. However, repeated injections are needed to achieve an observable effect^[50]. To avoid multiple injections, cell-based delivery of neurotrophic factors was proposed. A phase-I clinical trial for glaucoma (NCT01408472; <http://clinicaltrials.gov/>) using genetically modified CNTF-secreting retinal pigment epithelial cells (NT-501 CNTF implant) has already been launched—the outcomes have not been reported yet. Since MSCs can produce neurotrophic factors, including BDNF, CNTF, GDNF and basic fibroblast growth factor (bFGF), without the requirement of genetic modification, MSC transplantation has been suggested as a potential reservoir for neurotrophic factor secretion^[51]. Bone marrow-derived MSC transplantation increases RGC survival in a model of transient ischemia followed by reperfusion^[52], and reduces RGC loss in ocular hypertension models^[53,54]. Similarly, transplantation of human umbilical cord blood MSCs promotes RGC survival in an optic nerve crush model even after 7 d of injury^[55]. In addition, intracranial human umbilical cord blood MSC transplantation at the site of optic tract transection also protects RGCs and induces axonal regeneration^[56]. The neuroprotective effect of MSCs on RGC survival has clearly been proven, and the first clinical trial using bone marrow-derived MSCs on glaucoma in Florida (Stem Cell Ophthalmology Treatment Study (SCOTS)) has just started in August 2013 (NCT01920867; <http://clinicaltrials.gov/>; Table 2). This study will be complete in 2017.

RETINITIS PIGMENTOSA AND AGE-RELATED MACULAR DEGENERATION

Retinitis pigmentosa (RP) is characterized by a classic pattern of difficulties in dark adaptation and night blindness in adolescence, loss of mid-peripheral visual field in young adulthood and central vision later in life. These are due to the severe attenuation of rod and cone photoreceptors^[57]. RP is one of the hereditary degenerative diseases, affecting 1 in 4000 individuals. Age-related

Table 2 Registered clinical trials on mesenchymal stem cells for retinal diseases

Identifier	Country	Status	Study	Phase of trial	Estimated number of patients	Estimated trial end	Disease
NCT01531348	Thailand	Enrolling by invitation	Feasibility and safety of adult human bone marrow-derived mesenchymal stem cells by intravitreal injection in patients with retinitis pigmentosa	Phase 1	10	2014	Retinitis pigmentosa
NCT01914913	India	Not yet recruiting	Clinical study to evaluate safety and efficacy of stem cell therapy in retinitis pigmentosa	Phase 1/2	15	2015	Retinitis pigmentosa
NCT01920867	United States	Recruiting	Stem cell ophthalmology treatment study		300	2017	Glaucoma, retinitis pigmentosa, age-related macular degeneration

Information obtained from <http://clinicaltrials.gov/>.

macular degeneration (AMD) is the leading cause of irreversible blindness in people aged 50 years or above in the developed world^[58]. It influences the central portion of the retina (the macula). Early AMD is characterized by drusen (pale yellowish lesions), or by hyperpigmentation and hypopigmentation of retinal pigment epithelium in the macula. Late AMD is divided into the “non-exudative” and “exudative” forms. The non-exudative form (geographic atrophy) starts with a sharply demarcated round or oval hypopigmented spot in which large choroidal vessels are visible, whereas the exudative form, characterized by choroidal neovascularization, is the detachment of the neuroretina or RPE from Bruch’s membrane by serous or hemorrhagic fluid^[59,60].

Both RP and AMD involve photoreceptor cell death. MSC research studies targeting this common pathology can be divided into two categories: first, cell replacement-based studies aim to generate photoreceptor cells from different sources of MSCs. MSCs from the trabecular meshwork as well as the conjunctiva have been used to produce photoreceptor-like cells *in vitro*^[61,62]. Interestingly, subretinal injection of MSCs has also been reported to induce differentiation into photoreceptor cells in a sodium iodate-induced retinal degeneration rat model^[63]. Second, studies based on paracrine effects hypothesize that MSCs can secrete neurotrophic factors to protect against photoreceptor degeneration in different animal models. Transplantation of bone marrow-derived MSCs can rescue photoreceptor cells of the dystrophic retina in the rhodopsin knockout mouse model^[64]. Moreover, intravenous injection of bone marrow-derived MSCs rescue photoreceptor cells as well as visual function in the Royal College of Surgeons rat model^[65]. For AMD, beside photoreceptor cell loss, retinal pigment epithelial (RPE) cells are also affected. Adipose tissue-derived MSCs can be induced to an RPE phenotype^[66]. In addition, adipose tissue-derived MSCs rescue mitomycin C-treated RPE cell lines (ARPE19) from death in culture^[67]. Furthermore, subretinal injected MSCs adopt RPE morphology and preserve the retinal layer integrity in the sodium iodate-induced retinal degeneration rat model^[68].

To date, there are three ongoing registered clinical trials using MSCs on RP (Table 2). The first clinical trial

aims to determine the feasibility and safety of human adult bone marrow-derived MSCs by intravitreal injection in patients with RP in Thailand (NCT01531348; <http://clinicaltrials.gov/>). The second clinical trial is the Stem Cell Ophthalmology Treatment Study (SCOTS) in Florida (NCT01920867; <http://clinicaltrials.gov/>) proposed to use autologous bone marrow-derived MSCs by different means of injection (retrobulbar, subtenon, intravitreal, intraocular, subretinal and intravenous). The third clinical trial is a Phase-1/2 open labeled study done in India to evaluate the safety and efficacy of bone marrow-derived MSCs in RP (NCT01914913; <http://clinicaltrials.gov/>). For AMD, there is only one registered clinical trial using bone marrow-derived MSCs (Table 2), the Stem Cell Ophthalmology Treatment Study (SCOTS) in Florida (NCT01920867; <http://clinicaltrials.gov/>). Results from these studies have not been reported yet.

CONCLUSION

MSCs have been discovered for more than 20 years^[69], and have been found all over the body. MSCs can be conveniently obtained from different accessible tissues: bone marrow, blood, and adipose and dental tissue. They can also be easily expanded in standard culture conditions. In addition to the above mentioned characteristics, MSCs demonstrate neuroprotective effects, immunomodulatory properties and self-migratory activity, making them an attractive therapeutic tool. In recent years, MSC research has already begun the transition from preclinical experiments to human clinical trials. There are currently more than 60 MSC clinical trials dealing with CNS disorders and three clinical trials on retinal diseases. Although transient rash, self-limiting bacterial infections or fever might occur in some patients after MSC transplantation, serious adverse events have never been observed. This can foresee that MSC transplantation will become routine clinical practice for disease treatment in the near future. However, there are critical challenges still needed to be conquered before MSC therapy can be adopted in daily clinical practice. These include: (1) poor MSC retention *in vivo*; (2) poor MSC engraftment, viability and function *in vivo*; (3) unclear mechanisms of action; and (4) lack

of standardized trials^[70]. Moreover, few studies showed the contradictory results of MSC immunomodulatory properties. This might be explained by the heterogeneous MSC population. TLR4-primed human MSCs (MSC1) mostly secrete pro-inflammatory cytokines (IL-6, IL-8) while TLR3-primed human MSCs (MSC2) express mostly immunosuppressive mediators (IL-10, IDO, TSG-6)^[71]. Addition of fewer MSCs (10-1000) would lead to a less consistent suppression or a marked lymphocyte proliferation in culture, whereas addition of 10000-40000 MSCs have an inhibitory effect^[72]. Besides, there are uncertainties that must be answered. What is the optimal cell number for transplantation? Which MSC types are optimal for regenerative medicine? When is the optimal stage to receive MSC therapy? Which transplantation route is suitable for each individual CNS disorder? Further research is needed to understand the mechanisms elicited by stem cells in regenerating damaged tissues after transplantation.

REFERENCES

- 1 **Wagers AJ**, Weissman IL. Plasticity of adult stem cells. *Cell* 2004; **116**: 639-648 [PMID: 15006347 DOI: 10.1016/S0092-8674(04)00208-9]
- 2 **Horwitz EM**, Le Blanc K, Dominici M, Mueller I, Slaper-Cortenbach I, Marini FC, Deans RJ, Krause DS, Keating A. Clarification of the nomenclature for MSC: The International Society for Cellular Therapy position statement. *Cytotherapy* 2005; **7**: 393-395 [PMID: 16236628 DOI: 10.1080/14653240500319234]
- 3 **Ding DC**, Shyu WC, Lin SZ. Mesenchymal stem cells. *Cell Transplant* 2011; **20**: 5-14 [PMID: 21396235 DOI: 10.3727/096368910X]
- 4 **Dominici M**, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, Deans R, Keating A, Prockop DJ, Horwitz E. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006; **8**: 315-317 [PMID: 16923606 DOI: 10.1080/14653240600855905]
- 5 **Tárnok A**, Ulrich H, Bocsi J. Phenotypes of stem cells from diverse origin. *Cytometry A* 2010; **77**: 6-10 [PMID: 20024907 DOI: 10.1002/cyto.a.20844]
- 6 **Prockop DJ**. Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 1997; **276**: 71-74 [PMID: 9082988 DOI: 10.1126/science.276.5309.71]
- 7 **Pittenger MF**, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; **284**: 143-147 [PMID: 10102814 DOI: 10.1126/science.284.5411.143]
- 8 **Siniscalco D**, Giordano C, Galderisi U, Luongo L, Alessio N, Di Bernardo G, de Novellis V, Rossi F, Maione S. Intra-brain microinjection of human mesenchymal stem cells decreases allodynia in neuropathic mice. *Cell Mol Life Sci* 2010; **67**: 655-669 [PMID: 19937263 DOI: 10.1007/s00018-009-0202-4]
- 9 **Chen PM**, Yen ML, Liu KJ, Sytwu HK, Yen BL. Immunomodulatory properties of human adult and fetal multipotent mesenchymal stem cells. *J Biomed Sci* 2011; **18**: 49 [PMID: 21762539 DOI: 10.1186/1423-0127-18-49]
- 10 **Amado LC**, Saliaris AP, Schuleri KH, St John M, Xie JS, Cattaneo S, Durand DJ, Fitton T, Kuang JQ, Stewart G, Lehrke S, Baumgartner WW, Martin BJ, Heldman AW, Hare JM. Cardiac repair with intramyocardial injection of allogeneic mesenchymal stem cells after myocardial infarction. *Proc Natl Acad Sci USA* 2005; **102**: 11474-11479 [PMID: 16061805 DOI: 10.1073/pnas.0504388102]
- 11 **Kawanabe N**, Murata S, Murakami K, Ishihara Y, Hayano S, Kurosaka H, Kamioka H, Takano-Yamamoto T, Yamashiro T. Isolation of multipotent stem cells in human periodontal ligament using stage-specific embryonic antigen-4. *Differentiation* 2010; **79**: 74-83 [PMID: 19945209 DOI: 10.1016/j.diff.2009.10.005]
- 12 **Zarceczny A**, Caulfield T. Emerging ethical, legal and social issues associated with stem cell research & amp; and the current role of the moral status of the embryo. *Stem Cell Rev* 2009; **5**: 96-101 [PMID: 19521800 DOI: 10.1007/s12015-009-9062-4]
- 13 **Noseworthy JH**, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. *N Engl J Med* 2000; **343**: 938-952 [PMID: 11006371 DOI: 10.1056/NEJM200009283431307]
- 14 **Frohman EM**, Racke MK, Raine CS. Multiple sclerosis--the plaque and its pathogenesis. *N Engl J Med* 2006; **354**: 942-955 [PMID: 16510748 DOI: 10.1056/NEJMra052130]
- 15 **Bielekova B**, Goodwin B, Richert N, Cortese I, Kondo T, Afshar G, Gran B, Eaton J, Antel J, Frank JA, McFarland HF, Martin R. Encephalitogenic potential of the myelin basic protein peptide (amino acids 83-99) in multiple sclerosis: results of a phase II clinical trial with an altered peptide ligand. *Nat Med* 2000; **6**: 1167-1175 [PMID: 11017150 DOI: 10.1038/80516]
- 16 **Goodin DS**, Frohman EM, Garmany GP, Halper J, Likosky WH, Lublin FD, Silberberg DH, Stuart WH, van den Noort S. Disease modifying therapies in multiple sclerosis: report of the Therapeutics and Technology Assessment Subcommittee of the American Academy of Neurology and the MS Council for Clinical Practice Guidelines. *Neurology* 2002; **58**: 169-178 [PMID: 11805241 DOI: 10.1212/WNL.58.2.169]
- 17 **Mix E**, Meyer-Rienecker H, Hartung HP, Zettl UK. Animal models of multiple sclerosis--potentials and limitations. *Prog Neurobiol* 2010; **92**: 386-404 [PMID: 20558237 DOI: 10.1016/j.pneurobio.2010.06.005]
- 18 **Carrithers MD**. Current immunotherapy of multiple sclerosis and future challenges: relevance of immune-mediated repair. *Curr Pharm Biotechnol* 2012; **13**: 1409-1417 [PMID: 22339217 DOI: 10.2174/138920112800784781]
- 19 **Auletta JJ**, Bartholomew AM, Maziarz RT, Deans RJ, Miller RH, Lazarus HM, Cohen JA. The potential of mesenchymal stromal cells as a novel cellular therapy for multiple sclerosis. *Immunotherapy* 2012; **4**: 529-547 [PMID: 22642335 DOI: 10.2217/imt.12.41]
- 20 **Bai L**, Lennon DP, Eaton V, Maier K, Caplan AI, Miller SD, Miller RH. Human bone marrow-derived mesenchymal stem cells induce Th2-polarized immune response and promote endogenous repair in animal models of multiple sclerosis. *Glia* 2009; **57**: 1192-1203 [PMID: 19191336 DOI: 10.1002/glia.20841]
- 21 **Karussis D**, Karageorgiou C, Vakhnin-Dembinsky A, Gowda-Kurkalli B, Gomori JM, Kassis I, Bulte JW, Petrou P, Ben-Hur T, Abramsky O, Slavin S. Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis. *Arch Neurol* 2010; **67**: 1187-1194 [PMID: 20937945 DOI: 10.1001/archneurol.2010.248]
- 22 **Connick P**, Kolappan M, Patani R, Scott MA, Crawley C, He XL, Richardson K, Barber K, Webber DJ, Wheeler-Kingshott CA, Tozer DJ, Samson RS, Thomas DL, Du MQ, Luan SL, Michell AW, Altmann DR, Thompson AJ, Miller DH, Compston A, Chandran S. The mesenchymal stem cells in multiple sclerosis (MSCIMS) trial protocol and baseline cohort characteristics: an open-label pre-test: post-test study with blinded outcome assessments. *Trials* 2011; **12**: 62 [PMID: 21366911 DOI: 10.1186/1745-6215-12-62]
- 23 **Connick P**, Kolappan M, Crawley C, Webber DJ, Patani R, Michell AW, Du MQ, Luan SL, Altmann DR, Thompson AJ, Compston A, Scott MA, Miller DH, Chandran S. Autologous mesenchymal stem cells for the treatment of secondary pro-

- gressive multiple sclerosis: an open-label phase 2a proof-of-concept study. *Lancet Neurol* 2012; **11**: 150-156 [PMID: 22236384 DOI: 10.1016/S1474-4422(11)70305-2]
- 24 **Harkey HL**, White EA, Tibbs RE, Haines DE. A clinician's view of spinal cord injury. *Anat Rec B New Anat* 2003; **271**: 41-48 [PMID: 12619085 DOI: 10.1002/ar.b.10012]
- 25 **Furlan JC**, Sakakibara BM, Miller WC, Krassioukov AV. Global incidence and prevalence of traumatic spinal cord injury. *Can J Neurol Sci* 2013; **40**: 456-464 [PMID: 23786727]
- 26 **Forostyak S**, Jendelova P, Sykova E. The role of mesenchymal stromal cells in spinal cord injury, regenerative medicine and possible clinical applications. *Biochimie* 2013; **95**: 2257-2270 [PMID: 23994163 DOI: 10.1016/j.biochi.2013.08.004]
- 27 **Hulsebosch CE**. Recent advances in pathophysiology and treatment of spinal cord injury. *Adv Physiol Educ* 2002; **26**: 238-255 [PMID: 12443996]
- 28 **Bracken MB**, Shepard MJ, Collins WF, Holford TR, Young W, Baskin DS, Eisenberg HM, Flamm E, Leo-Summers L, Maroon J. A randomized, controlled trial of methylprednisolone or naloxone in the treatment of acute spinal-cord injury. Results of the Second National Acute Spinal Cord Injury Study. *N Engl J Med* 1990; **322**: 1405-1411 [PMID: 2278545 DOI: 10.1056/NEJM199005173222001]
- 29 **Zurita M**, Vaquero J, Bonilla C, Santos M, De Haro J, Oya S, Aguayo C. Functional recovery of chronic paraplegic pigs after autologous transplantation of bone marrow stromal cells. *Transplantation* 2008; **86**: 845-853 [PMID: 18813110 DOI: 10.1097/TP.0b013e318186198f]
- 30 **Hejcl A**, Sedý J, Kapcalová M, Toro DA, Amemori T, Lesný P, Likavcanová-Masínová K, Krumbholcová E, Prádný M, Michálek J, Burian M, Hájek M, Jendelová P, Syková E. HPMA-RGD hydrogels seeded with mesenchymal stem cells improve functional outcome in chronic spinal cord injury. *Stem Cells Dev* 2010; **19**: 1535-1546 [PMID: 20053128 DOI: 10.1089/scd.2009.0378]
- 31 **Gu W**, Zhang F, Xue Q, Ma Z, Lu P, Yu B. Transplantation of bone marrow mesenchymal stem cells reduces lesion volume and induces axonal regrowth of injured spinal cord. *Neuropathology* 2010; **30**: 205-217 [PMID: 19845866 DOI: 10.1111/j.1440-1789.2009.01063.x]
- 32 **Hejcl A**, Jendelová P, Syková E. Experimental reconstruction of the injured spinal cord. *Adv Tech Stand Neurosurg* 2011; **37**: 65-95 [PMID: 21997741 DOI: 10.1007/978-3-7091-0673-0_3]
- 33 **Ra JC**, Shin IS, Kim SH, Kang SK, Kang BC, Lee HY, Kim YJ, Jo JY, Yoon EJ, Choi HJ, Kwon E. Safety of intravenous infusion of human adipose tissue-derived mesenchymal stem cells in animals and humans. *Stem Cells Dev* 2011; **20**: 1297-1308 [PMID: 21303266 DOI: 10.1089/scd.2010.0466]
- 34 **Lauritsen MB**. Autism spectrum disorders. *Eur Child Adolesc Psychiatry* 2013; **22** Suppl 1: S37-S42 [PMID: 23300017 DOI: 10.1007/s00787-012-0359-5]
- 35 **Thompson T**. Autism research and services for young children: history, progress and challenges. *J Appl Res Intellect Disabil* 2013; **26**: 81-107 [PMID: 23404617 DOI: 10.1111/jar.12021]
- 36 **Levy SE**, Mandell DS, Schultz RT. Autism. *Lancet* 2009; **374**: 1627-1638 [PMID: 19819542 DOI: 10.1016/S0140-6736(09)61376-3]
- 37 **Johnson NL**, Rodriguez D. Children with autism spectrum disorder at a pediatric hospital: a systematic review of the literature. *Pediatr Nurs* 2013; **39**: 131-141 [PMID: 23926752]
- 38 **Williams K**, Wheeler DM, Silove N, Hazell P. Selective serotonin reuptake inhibitors (SSRIs) for autism spectrum disorders (ASD). *Cochrane Database Syst Rev* 2010; **(8)**: CD004677 [PMID: 20687077]
- 39 **McCracken JT**. Safety issues with drug therapies for autism spectrum disorders. *J Clin Psychiatry* 2005; **66** Suppl 10: 32-37 [PMID: 16401148]
- 40 **Nevels RM**, Dehon EE, Alexander K, Gontkovsky ST. Psychopharmacology of aggression in children and adolescents with primary neuropsychiatric disorders: a review of current and potentially promising treatment options. *Exp Clin Psychopharmacol* 2010; **18**: 184-201 [PMID: 20384430 DOI: 10.1037/a0018059]
- 41 **Ichim TE**, Solano F, Glenn E, Morales F, Smith L, Zabrecky G, Riordan NH. Stem cell therapy for autism. *J Transl Med* 2007; **5**: 30 [PMID: 17597540 DOI: 10.1186/1479-5876-5-30]
- 42 **Wada N**, Gronthos S, Bartold PM. Immunomodulatory effects of stem cells. *Periodontol 2000* 2013; **63**: 198-216 [PMID: 23931061 DOI: 10.1111/prd.12024]
- 43 **Siniscalco D**, Sapone A, Cirillo A, Giordano C, Maione S, Antonucci N. Autism spectrum disorders: is mesenchymal stem cell personalized therapy the future? *J Biomed Biotechnol* 2012; **2012**: 480289 [PMID: 22496609]
- 44 **Lv YT**, Zhang Y, Liu M, Qiuwaxi JN, Ashwood P, Cho SC, Huan Y, Ge RC, Chen XW, Wang ZJ, Kim BJ, Hu X. Transplantation of human cord blood mononuclear cells and umbilical cord-derived mesenchymal stem cells in autism. *J Transl Med* 2013; **11**: 196 [PMID: 23978163 DOI: 10.1186/1479-5876-11-196]
- 45 **Weinreb RN**, Khaw PT. Primary open-angle glaucoma. *Lancet* 2004; **363**: 1711-1720 [PMID: 15158634 DOI: 10.1016/S0140-6736(04)16257-0]
- 46 **Quigley HA**. Number of people with glaucoma worldwide. *Br J Ophthalmol* 1996; **80**: 389-393 [PMID: 8695555 DOI: 10.1136/bjo.80.5.389]
- 47 **Dietlein TS**, Hermann MM, Jordan JF. The medical and surgical treatment of glaucoma. *Dtsch Arztebl Int* 2009; **106**: 597-605; quiz 606 [PMID: 19890428]
- 48 **Almasieh M**, Wilson AM, Morquette B, Cueva Vargas JL, Di Polo A. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res* 2012; **31**: 152-181 [PMID: 22155051 DOI: 10.1016/j.preteyeres.2011.11.002]
- 49 **Johnson TV**, Bull ND, Martin KR. Neurotrophic factor delivery as a protective treatment for glaucoma. *Exp Eye Res* 2011; **93**: 196-203 [PMID: 20685205 DOI: 10.1016/j.exer.2010.05.016]
- 50 **Ko ML**, Hu DN, Ritch R, Sharma SC, Chen CF. Patterns of retinal ganglion cell survival after brain-derived neurotrophic factor administration in hypertensive eyes of rats. *Neurosci Lett* 2001; **305**: 139-142 [PMID: 11376903 DOI: 10.1016/S0304-3940(01)01830-4]
- 51 **Ng TK**, Lam DS, Cheung HS. Prospects of stem cells for retinal diseases. *Asia Pac J Ophthalmol* 2013; **2**: 57-63 [DOI: 10.1097/APO.0b013e31827e3e5d]
- 52 **Li N**, Li XR, Yuan JQ. Effects of bone-marrow mesenchymal stem cells transplanted into vitreous cavity of rat injured by ischemia/reperfusion. *Graefes Arch Clin Exp Ophthalmol* 2009; **247**: 503-514 [PMID: 19084985 DOI: 10.1007/s00417-008-1009-y]
- 53 **Yu S**, Tanabe T, Dezawa M, Ishikawa H, Yoshimura N. Effects of bone marrow stromal cell injection in an experimental glaucoma model. *Biochem Biophys Res Commun* 2006; **344**: 1071-1079 [PMID: 16643846 DOI: 10.1016/j.bbrc.2006.03.231]
- 54 **Johnson TV**, Bull ND, Hunt DP, Marina N, Tomarev SI, Martin KR. Neuroprotective effects of intravitreal mesenchymal stem cell transplantation in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2010; **51**: 2051-2059 [PMID: 19933193 DOI: 10.1167/iovs.09-4509]
- 55 **Zhao T**, Li Y, Tang L, Li Y, Fan F, Jiang B. Protective effects of human umbilical cord blood stem cell intravitreal transplantation against optic nerve injury in rats. *Graefes Arch Clin Exp Ophthalmol* 2011; **249**: 1021-1028 [PMID: 21360302 DOI: 10.1007/s00417-011-1635-7]
- 56 **Zwart I**, Hill AJ, Al-Allaf F, Shah M, Girdlestone J, Sanusi AB, Mehmet H, Navarrete R, Navarrete C, Jen LS. Umbilical cord blood mesenchymal stromal cells are neuroprotective and promote regeneration in a rat optic tract model. *Exp Neurol* 2009; **216**: 439-448 [PMID: 19320003 DOI: 10.1016/j.expneurol.2008.12.028]
- 57 **Hartong DT**, Berson EL, Dryja TP. Retinitis pigmentosa.

- Lancet* 2006; **368**: 1795-1809 [PMID: 17113430 DOI: 10.1016/S0140-6736(06)69740-7]
- 58 **Pascalini D**, Mariotti SP, Pokharel GP, Pararajasegaram R, Etya'ale D, Négrel AD, Resnikoff S. 2002 global update of available data on visual impairment: a compilation of population-based prevalence studies. *Ophthalmic Epidemiol* 2004; **11**: 67-115 [PMID: 15255026 DOI: 10.1076/opep.11.2.67.28158]
- 59 **Ng TK**, Yam GH, Chen WQ, Lee VY, Chen H, Chen LJ, Choy KW, Yang Z, Pang CP. Interactive expressions of HtrA1 and VEGF in human vitreous humors and fetal RPE cells. *Invest Ophthalmol Vis Sci* 2011; **52**: 3706-3712 [PMID: 21310902 DOI: 10.1167/iovs.10-6773]
- 60 **Ng TK**, Liang XY, Pang CP. HTRA1 in age-related macular degeneration. *Asia Pac J Ophthalmol* 2012; **1**: 51-63 [DOI: 10.1097/APO.0b013e31823e57fe]
- 61 **Nadri S**, Yazdani S, Arefian E, Gohari Z, Eslaminejad MB, Kazemi B, Soleimani M. Mesenchymal stem cells from trabecular meshwork become photoreceptor-like cells on amniotic membrane. *Neurosci Lett* 2013; **541**: 43-48 [PMID: 23403103 DOI: 10.1016/j.neulet.2012.12.055]
- 62 **Nadri S**, Kazemi B, Eslaminejad MB, Yazdani S, Soleimani M. High yield of cells committed to the photoreceptor-like cells from conjunctiva mesenchymal stem cells on nanofibrous scaffolds. *Mol Biol Rep* 2013; **40**: 3883-3890 [PMID: 23588957 DOI: 10.1007/s11033-012-2360-y]
- 63 **Huo DM**, Dong FT, Yu WH, Gao F. Differentiation of mesenchymal stem cell in the microenvironment of retinitis pigmentosa. *Int J Ophthalmol* 2010; **3**: 216-219 [PMID: 22553557]
- 64 **Arnhold S**, Absenger Y, Klein H, Addicks K, Schraermeyer U. Transplantation of bone marrow-derived mesenchymal stem cells rescue photoreceptor cells in the dystrophic retina of the rhodopsin knockout mouse. *Graefes Arch Clin Exp Ophthalmol* 2007; **245**: 414-422 [PMID: 16896916 DOI: 10.1007/s00417-006-0382-7]
- 65 **Wang S**, Lu B, Girman S, Duan J, McFarland T, Zhang QS, Grompe M, Adams G, Appukuttan B, Lund R. Non-invasive stem cell therapy in a rat model for retinal degeneration and vascular pathology. *PLoS One* 2010; **5**: e9200 [PMID: 20169166 DOI: 10.1371/journal.pone.0009200]
- 66 **Vossmerbaeumer U**, Ohnesorge S, Kuehl S, Haapalahti M, Kluter H, Jonas JB, Thierse HJ, Bieback K. Retinal pigment epithelial phenotype induced in human adipose tissue-derived mesenchymal stromal cells. *Cytotherapy* 2009; **11**: 177-188 [PMID: 19241195 DOI: 10.1080/14653240802714819]
- 67 **Singh AK**, Srivastava GK, García-Gutiérrez MT, Pastor JC. Adipose derived mesenchymal stem cells partially rescue mitomycin C treated ARPE19 cells from death in co-culture condition. *Histol Histopathol* 2013; **28**: 1577-1583 [PMID: 23719745]
- 68 **Guan Y**, Cui L, Qu Z, Lu L, Wang F, Wu Y, Zhang J, Gao F, Tian H, Xu L, Xu G, Li W, Jin Y, Xu GT. Subretinal transplantation of rat MSCs and erythropoietin gene modified rat MSCs for protecting and rescuing degenerative retina in rats. *Curr Mol Med* 2013; **13**: 1419-1431 [PMID: 23971737 DOI: 10.2174/15665240113139990071]
- 69 **Caplan AI**. Mesenchymal stem cells. *J Orthop Res* 1991; **9**: 641-650 [PMID: 1870029 DOI: 10.1002/jor.1100090504]
- 70 **Psaltis PJ**, Zannettino AC, Worthley SG, Gronthos S. Concise review: mesenchymal stromal cells: potential for cardiovascular repair. *Stem Cells* 2008; **26**: 2201-2210 [PMID: 18599808 DOI: 10.1634/stemcells.2008-0428]
- 71 **Waterman RS**, Tomchuck SL, Henkle SL, Betancourt AM. A new mesenchymal stem cell (MSC) paradigm: polarization into a pro-inflammatory MSC1 or an Immunosuppressive MSC2 phenotype. *PLoS One* 2010; **5**: e10088 [PMID: 20436665 DOI: 10.1371/journal.pone.0010088]
- 72 **Le Blanc K**, Tammik L, Sundberg B, Haynesworth SE, Ringdén O. Mesenchymal stem cells inhibit and stimulate mixed lymphocyte cultures and mitogenic responses independently of the major histocompatibility complex. *Scand J Immunol* 2003; **57**: 11-20 [PMID: 12542793 DOI: 10.1046/j.1365-3083.2003.01176.x]

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